



Regional Water Quality Issues: Algae and Associated Drinking Water Challenges

Workshop – August 2005

**A Cooperative Research and Implementation Program
Arizona State University (Tempe, AZ)
Milton Sommerfeld, Paul Westerhoff, Qiang Hu, John Crittenden,
Youngll Kim, Bo Song, Tom Dempster, Everett Shock, Panjai
Prapaipong, Larry Baker and Marisa Masles**

**Salt River Project
Central Arizona Project
City of Phoenix
City of Tempe
City of Peoria
City of Chandler
ASU NSF Water Quality Center**



Agenda

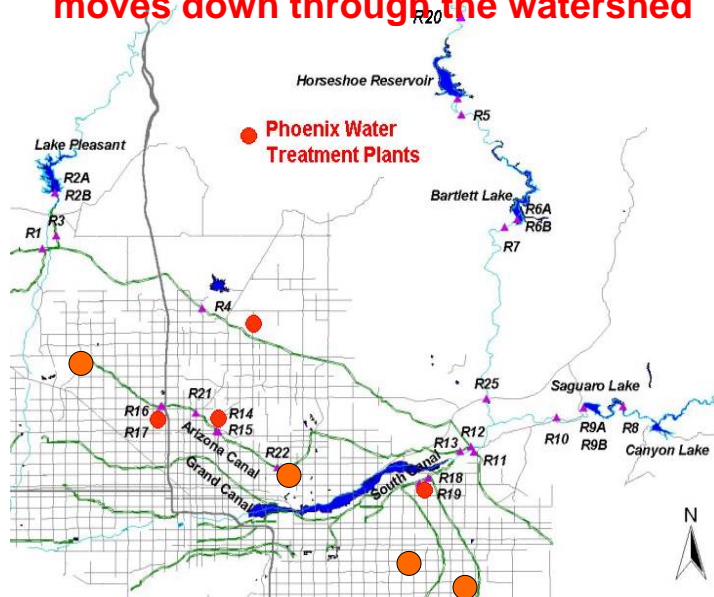
Purpose: Provide a forum to review and discuss on-going regional water quality issues, in particular algae-associated issues.

- 8:30 Introductions**
- 8:45 Overview of T&O issues for 2005**
- 9:15 Trace metals in the water supplies**
- 9:30 Fundamentals of GAC: T&O and DOC control**
- 10:00 Break**
- 10:15 Recent Progress on DNA-based probes for T&O and toxin producing algae**
- 10:40 Sonication for algae control**
- 10:50 Future directions & discussion**
- 11:00 Meeting adjournment**

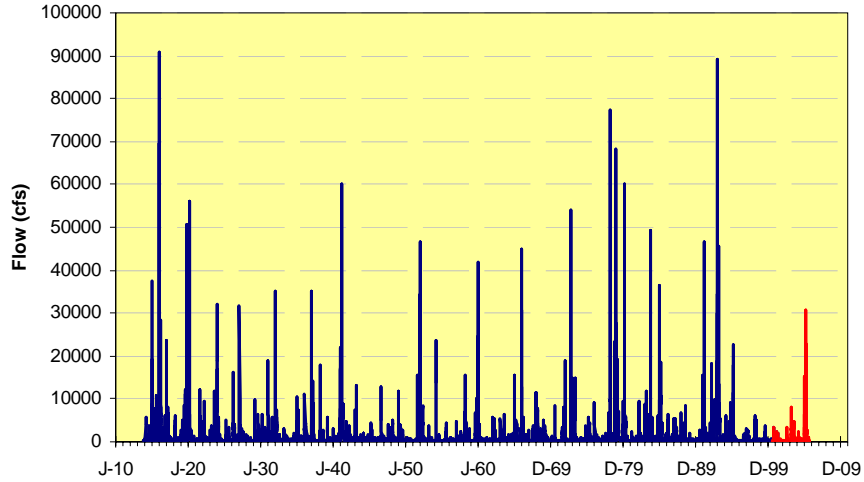
Overview of T&O issues for 2005

What is unique about 2005?

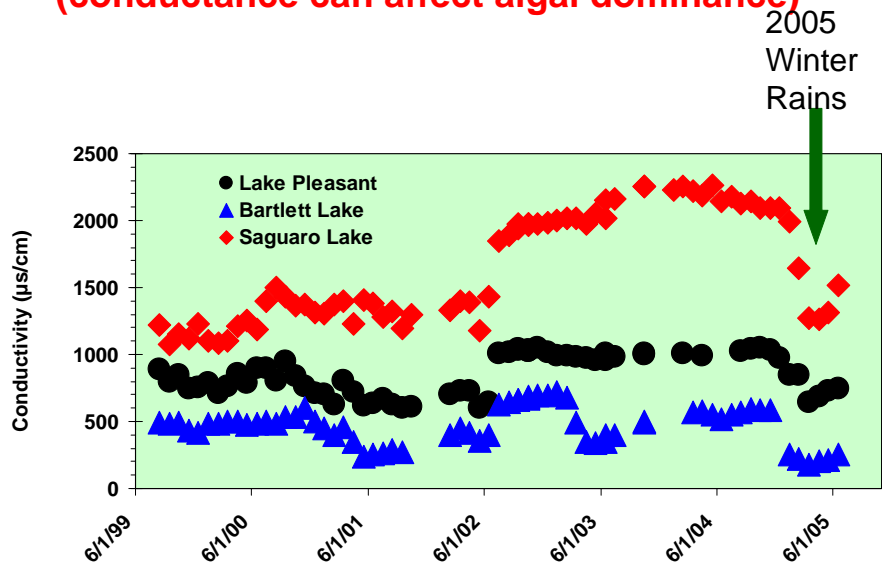
Workshop will present results as water moves down through the watershed



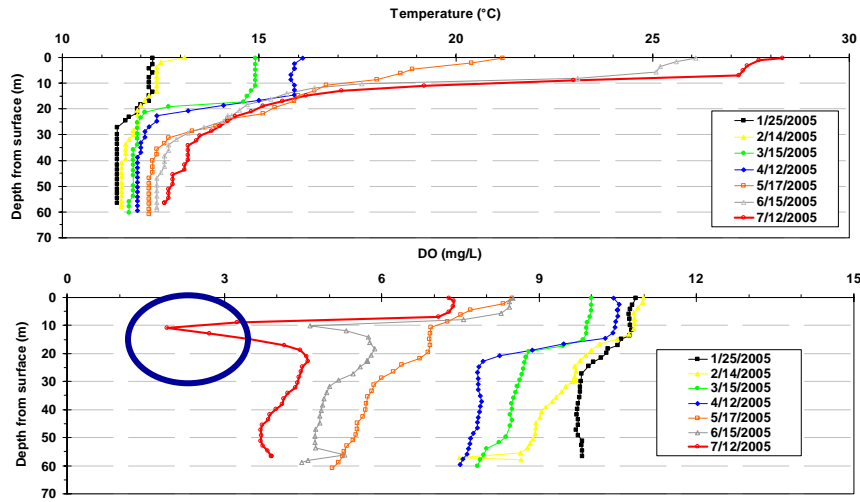
Salt River Above Roosevelt



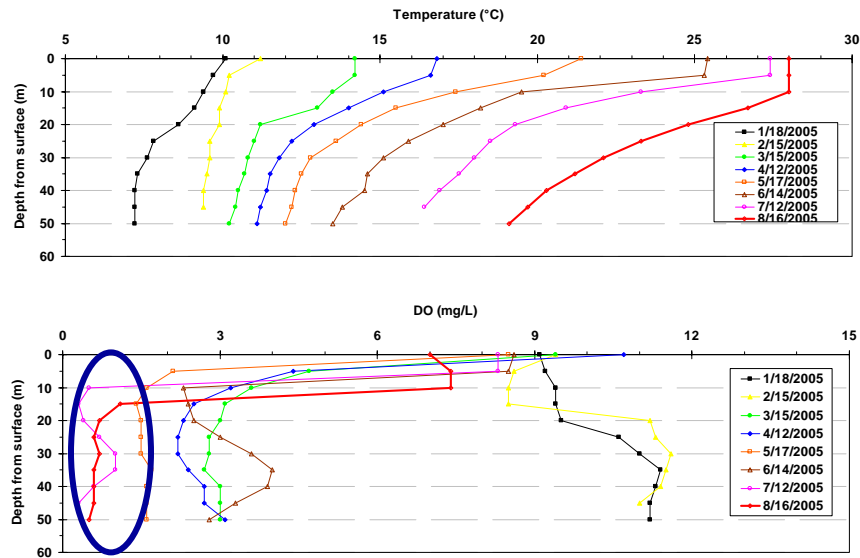
Hydrology Affects Water Quality (conductance can affect algal dominance)

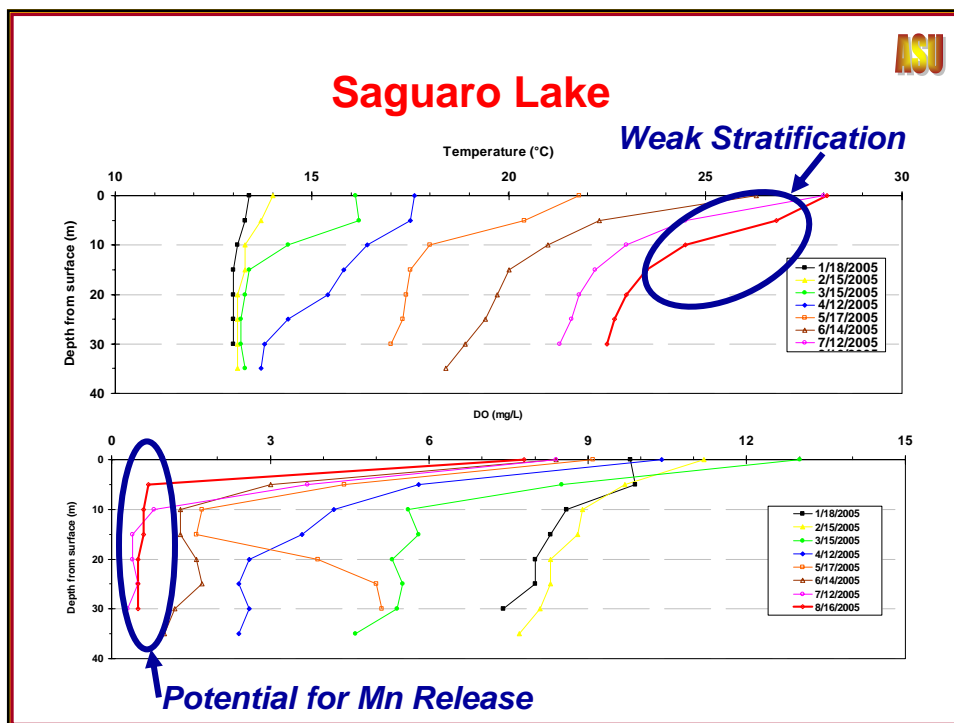
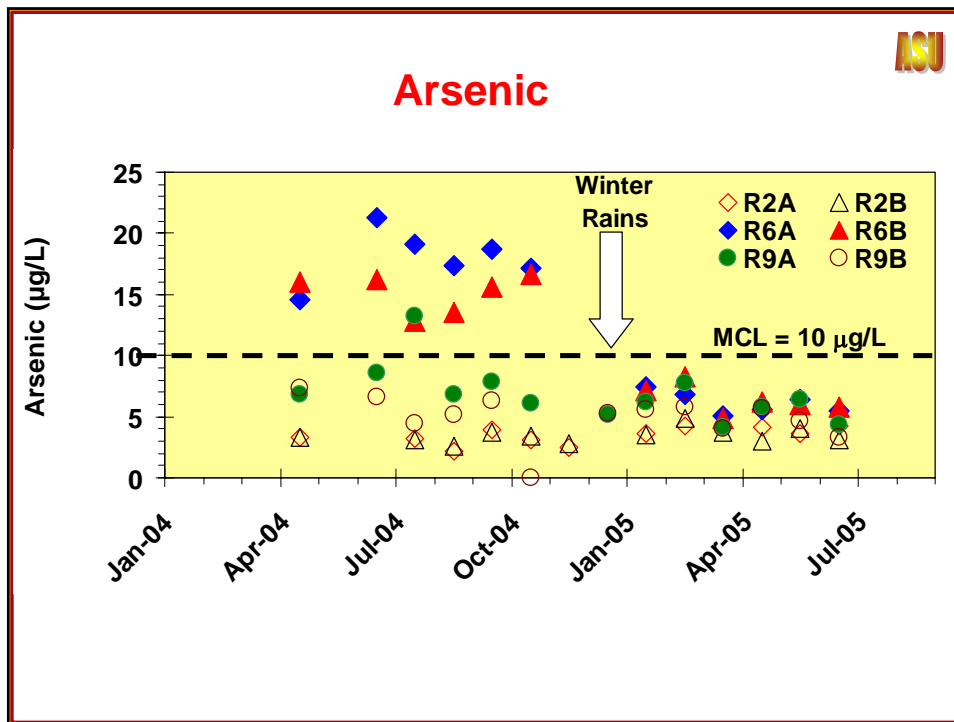


Lake Pleasant

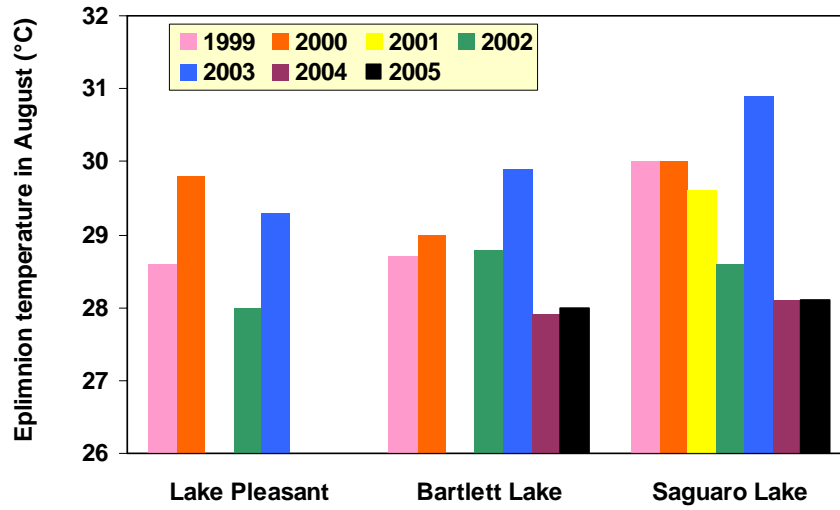


Bartlett Lake

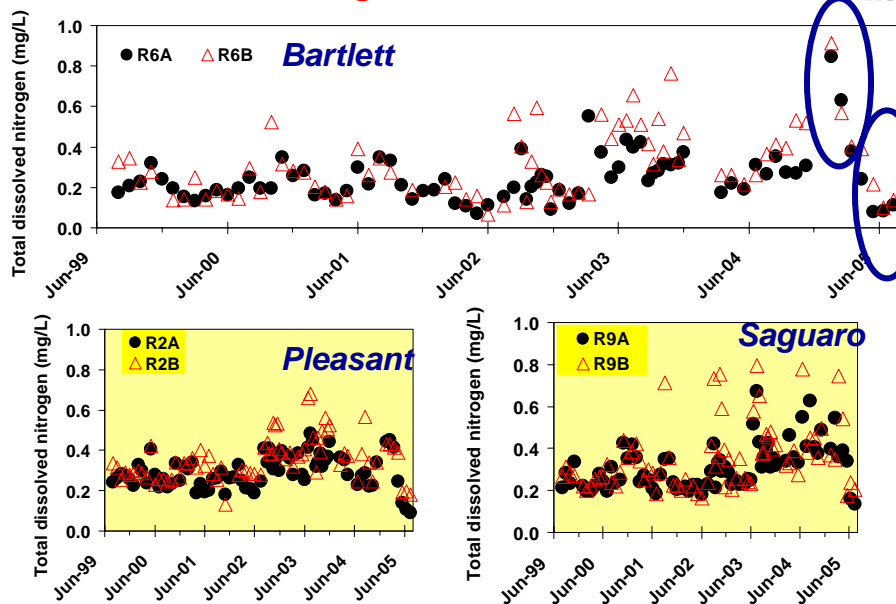


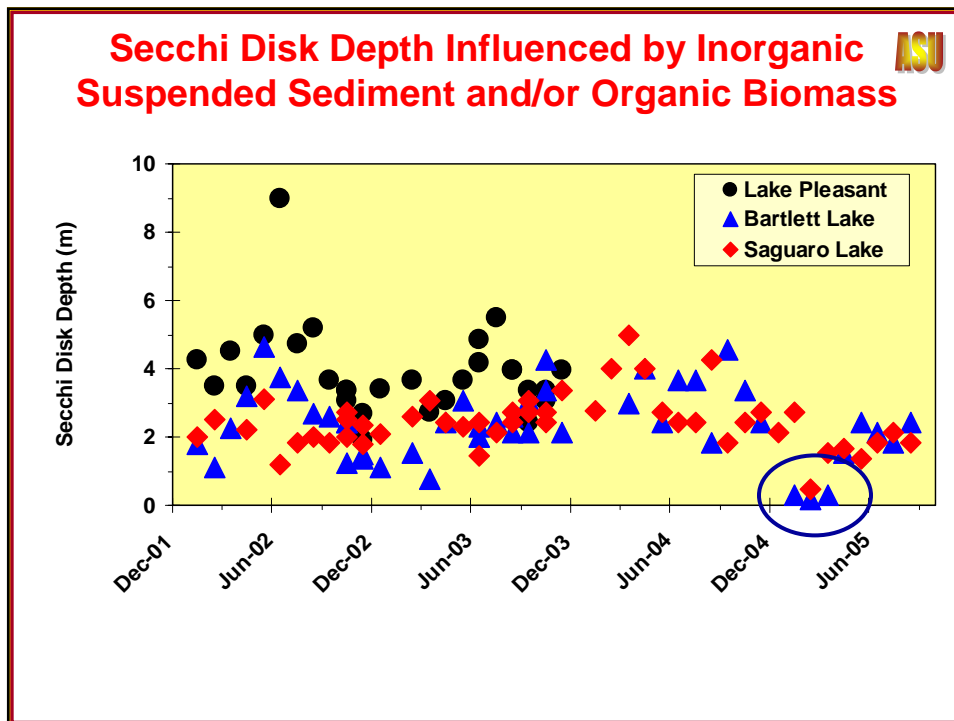
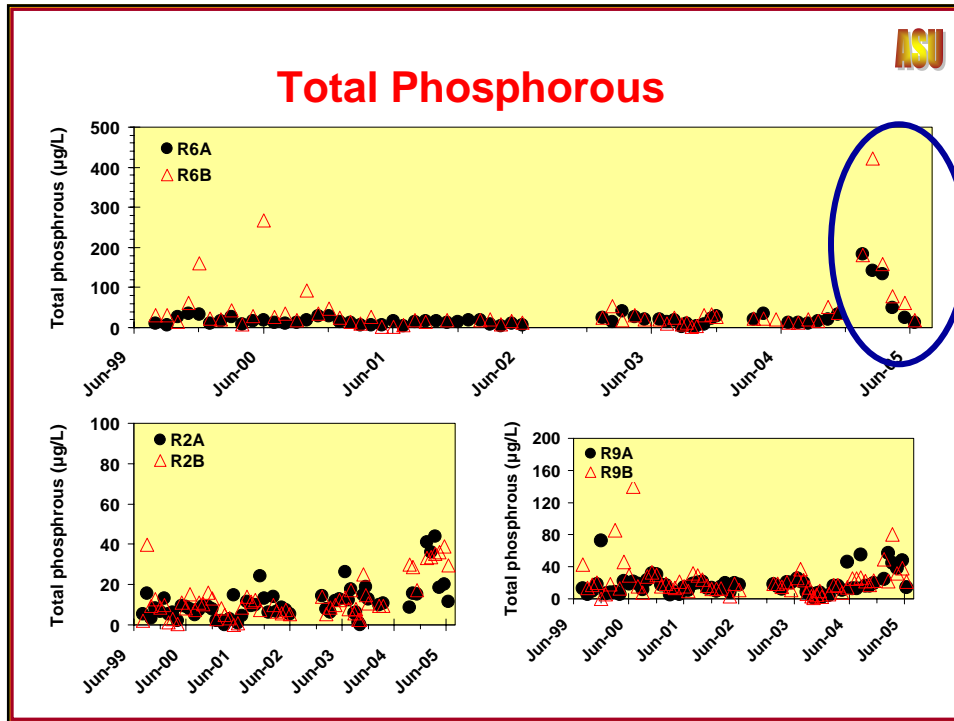


Summary of Annual August Temperatures

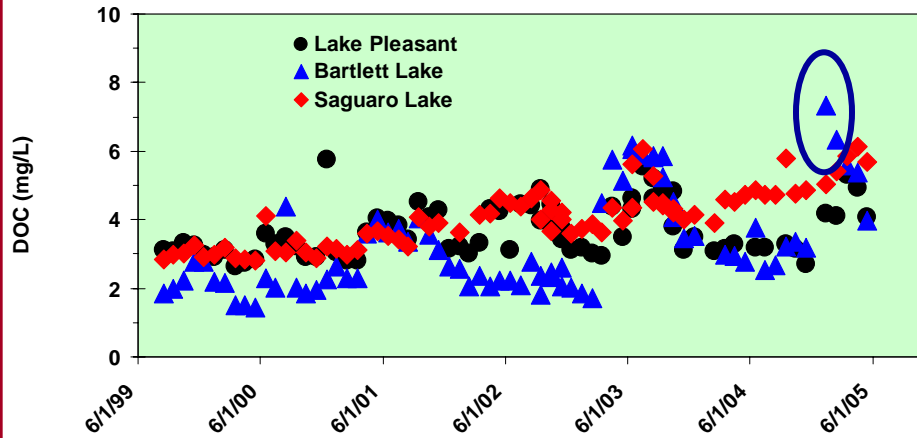


Dissolved Nitrogen Trends in Reservoirs

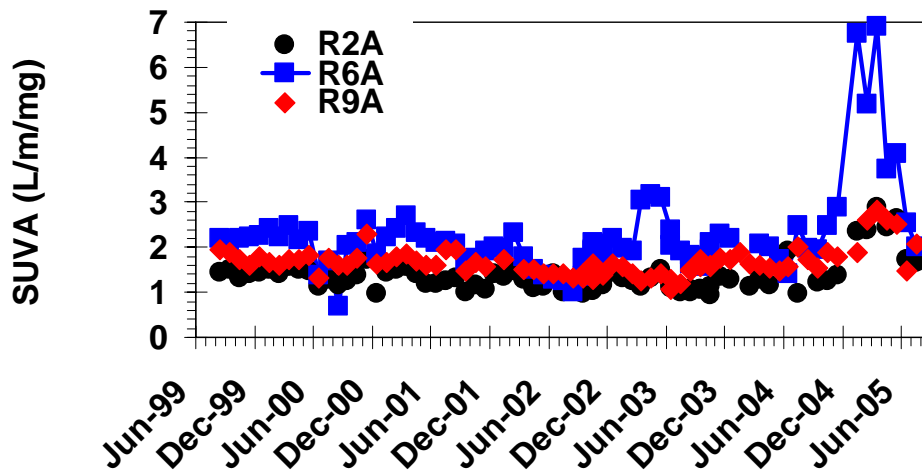




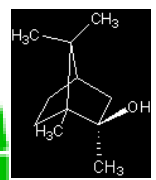
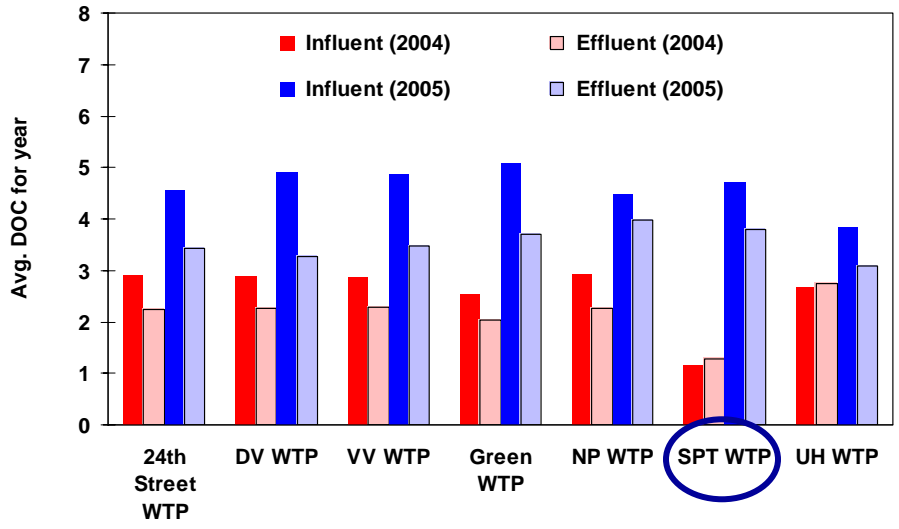
Up-stream reservoirs attenuate DOC



Specific UV Absorbance at 254 nm



DOC Removal by WTP

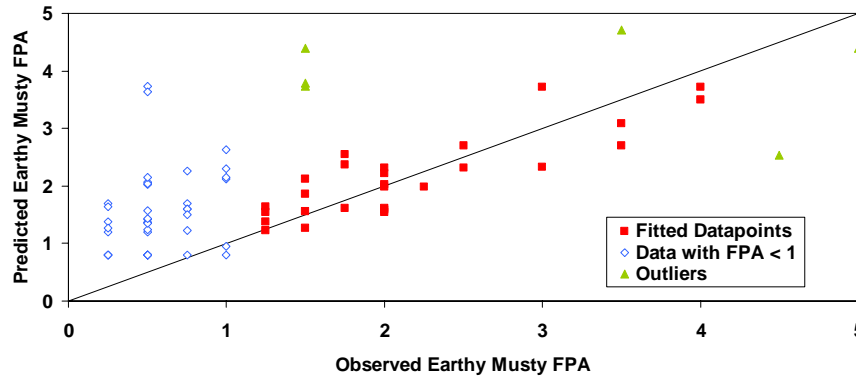


Predicting Expected FPA Intensity based upon GC/MS data



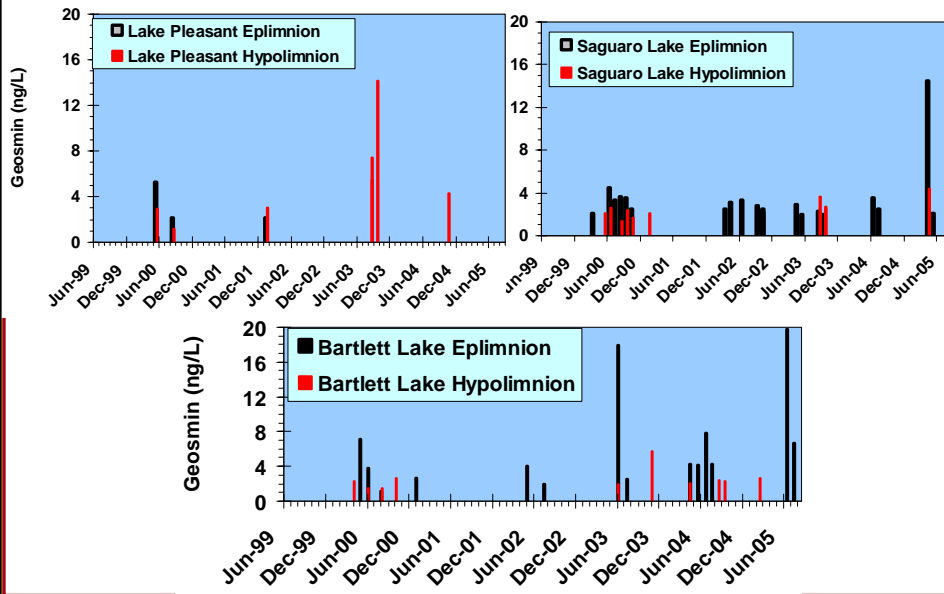
$$\text{Earthy Musty FPA Value} = 0.800 \cdot \text{MIB}^{0.396} \cdot \text{Geo}^{-0.110} \cdot \text{Cyclocitral}^{0.350}$$

$$R^2 = 0.728$$

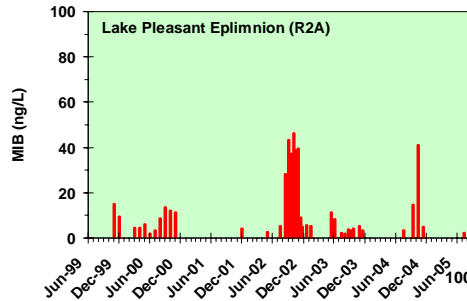


FPA= 2 when MIB=5, Geosmin=3, Cyclocitral=3 ng/L
FPA= 3 when MIB=8, Geosmin=8, Cyclocitral=8 ng/L

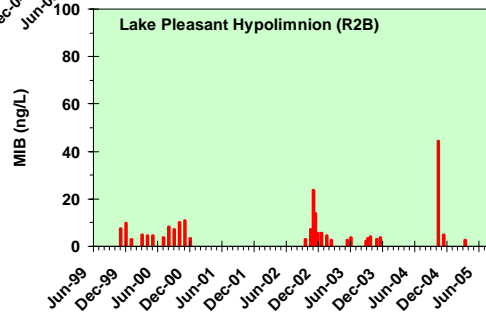
Geosmin Data



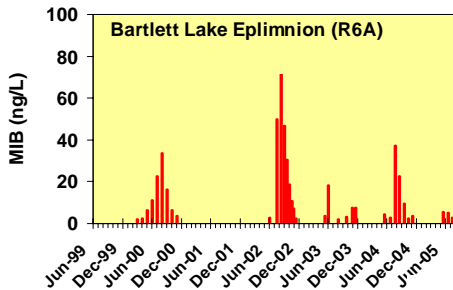
MIB Data – Lake Pleasant



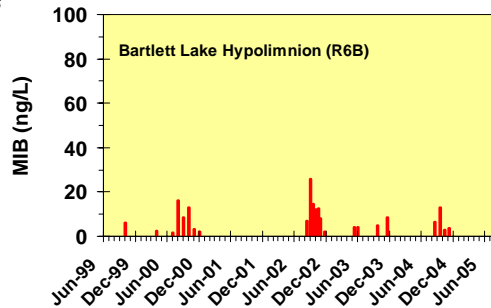
MIB is low in Lake Pleasant in 2005



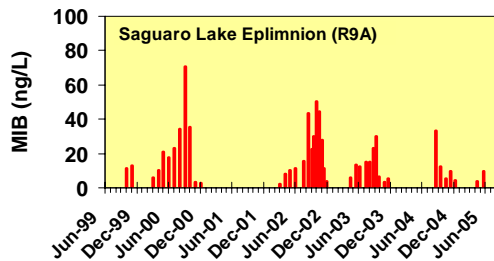
MIB Data – Bartlett Lake



MIB is low in Bartlett Reservoir in 2005



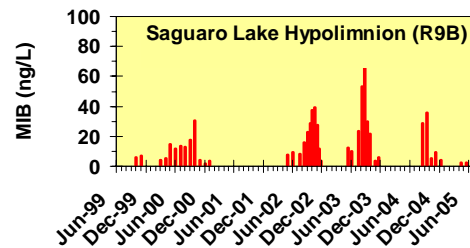
MIB Data – Saguaro Lake



MIB is low in Saguaro Lake in 2005

Conductance in reservoirs has been a general indicator for T&O

Blue-green algae prefer higher TDS



Baker MIB Production/Loss Mass Balance Model for Canals

MIB load leaving segment = MIB load entering segment + production within the segment – MIB lost via diversions

- For the simple case of one diversion, the model is:

$$[\text{MIB}]_l \cdot Q_l \cdot 10^{-6} = [\text{MIB}]_u \cdot Q_u \cdot 10^{-6} + k \cdot L - k \cdot L_d \cdot Q_{d,\text{out}} / Q_{d,\text{AZ}}$$

- Where

- ◆ $[\text{MIB}]_u$ = MIB concentration at upper end of segment
- ◆ Q_u = flow at upper end of segment, m³/day
- ◆ $[\text{MIB}]_l$ = MIB concentration at lower end of segment, ng/L
- ◆ Q_l = flow at lower end of segment
- ◆ k = MIB production rate (0th order), g/mile
- ◆ L = length of segment, miles
- ◆ L_d = length of segment from upper end down to diversion within a segment
- ◆ $Q_{d,\text{out}}$ = flow from diversion, m³/day
- ◆ $Q_{d,\text{AZ}}$ = flow in the Arizona Canal at the point of diversion

**Predict “k” values along Arizona Canal
for loss of MIB (g/mile-day)**



Canal Segment	8/4/03	8/25/03	10/23/03	7/6/04	9/28/04
Below X-connect to Pima Road	2	6.1	9.2	0.5	2.1
Pima Road to 24 th Street WTP	0.6	-1	1.7	3.7	1.7
24 th Street WTP to Deer Valley WTP	3.4	2.5	1.5	1	0.33
Deer Valley WTP to Greenway WTP	-1.6	-5.5	-4.4	-0.1	-5.1

MIB Production

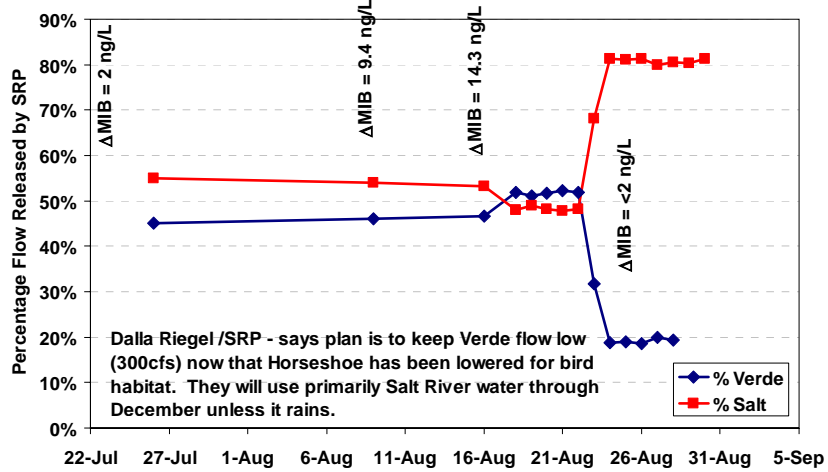
MIB Loss

**Recent Trends of In-situ MIB Production (Δ MIB in ng/L)
in Arizona Canal
(Geosmin and Cyclocitral follow similar patterns)**

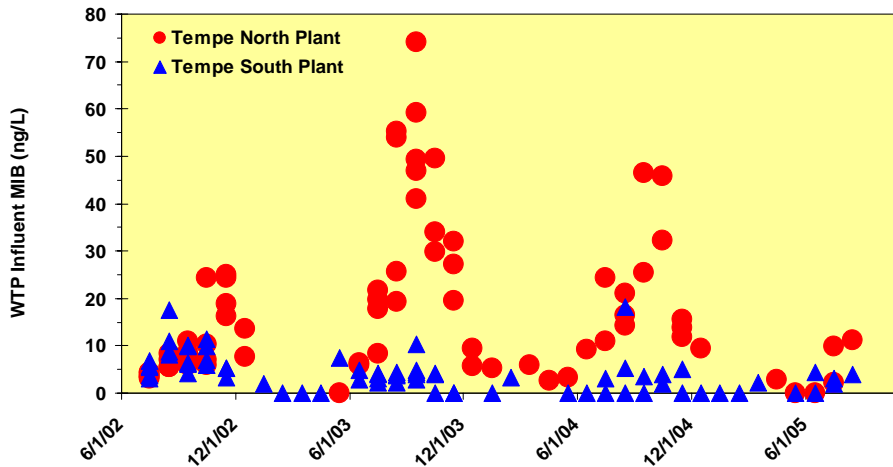


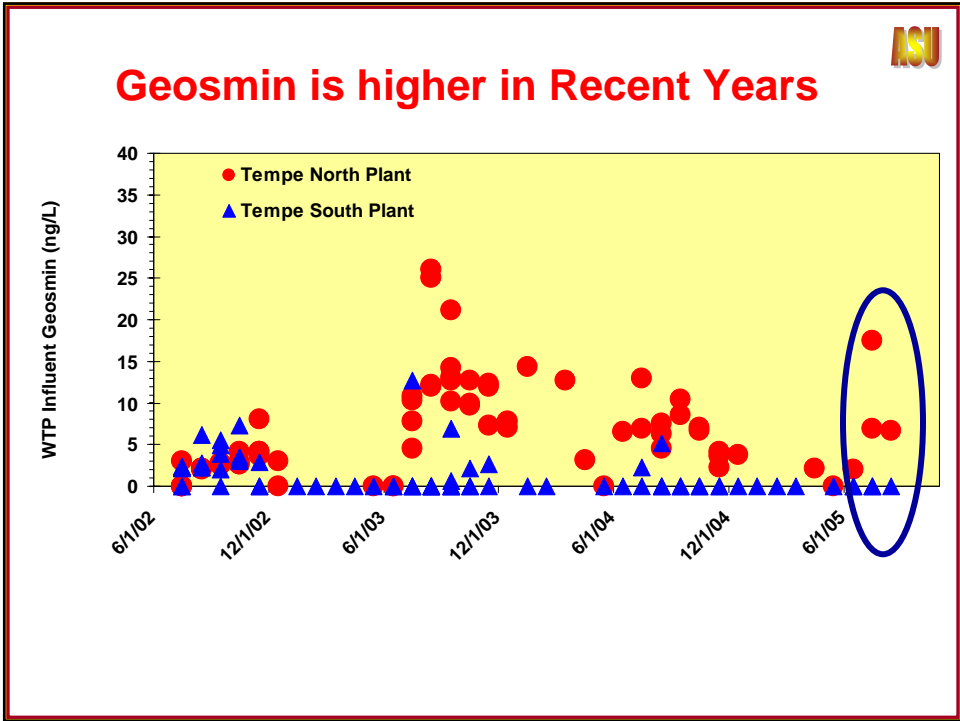
Canal Segment	6/28/05	7/12/05	7/26/05	8/16/05	8/25/05
MIB below X-connect	<2 ng/L	<2 ng/L	3.1 ng/L	4.8 ng/L	
Below X-connect to Pima Road	2.5	1.0	7.0	7.4	
Pima Road to 24 th Street WTP	1.2	~0	~0	3.3	~0
24 th Street WTP to Deer Valley WTP	-0.8	0.7	2.4	3.6	
Deer Valley WTP to Greenway WTP	--	1.5	-3.2	-4.1	

SRP Change in River Release affected MIB Production in the canals – suggests conductance effect



MIB levels higher in AZ Canal system compared against South Canal system





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Table 3 - Canal Sampling – August 30, 2005

System	Sample Description	MIB (ng/L)	Geosmin (ng/L)	Cyclocitral (ng/L)
CAP	Waddell Canal	<2.0	<2.0	<2.0
	Waddell Canal - CAP	<2.0	<2.0	<2.0
	Union Hills Inlet	<2.0	2.5	2.4
	CAP Canal at Cross-connect	2.0	3.9	3.5
AZ Canal	Salt River @ Blue Pt Bridge	18.2	3.0	2.8
	Verde River @ Beeline	21.8	2.1	3.7
	AZ Canal above CAP Cross-connect	14.8	2.4	<2.0
	AZ Canal below CAP Cross-connect	13.4	2.7	<2.0
	AZ Canal at Highway 87	14.3	3.8	2.5
	AZ Canal at Pima Rd.	15.8	4.4	3.3
	AZ Canal at 56th St.	13.2	3.9	2.9
	AZ Canal - Inlet to 24 th Street WTP	13.9	4.3	3.0
	AZ Canal - Central Avenue	16.9	4.9	3.0
	AZ Canal - Inlet to Deer Valley WTP	20.8	7.2	3.4
South and Tempe Canals	AZ Canal - Inlet to Greenway WTP	16.0	3.9	<2.0
	South Canal below CAP Cross-connect	16.2	2.6	<2.0
	South Canal at Val Vista WTP	17.4	3.0	<2.0
	Head of the Tempe Canal	15.2	2.6	<2.0
Canals	Tempe Canal - Inlet to Tempe's South Plant	8.8	<2.0	<2.0
	Chandler WTP – Inlet			

Trace Metals : Cycles, Transport and Urban Signatures

Panjai Prapaipong*, Brandon McLean,
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*GEOPIG, Department of Geological Science
Department of Chemistry and Biochemistry
Arizona State University
panjai@asu.edu*

GEOPIG Analytical Facility



**High Resolution ICP-MS
(minor & trace elements)**



**Ion Chromatography
(major ions)**

Others: MS for stable isotopes & isotope ratio, quadrupole MS,
GC, GC-MS, microwave digester

<http://geopig.asu.edu>

Projects: Environmental Biogeochemistry

- Human-induced biogeochemical cycles of metals
- Chemical footprint of cities, using the Phoenix metropolitan area as a guide
- Micronutrient transport in rivers in response to climate forcing

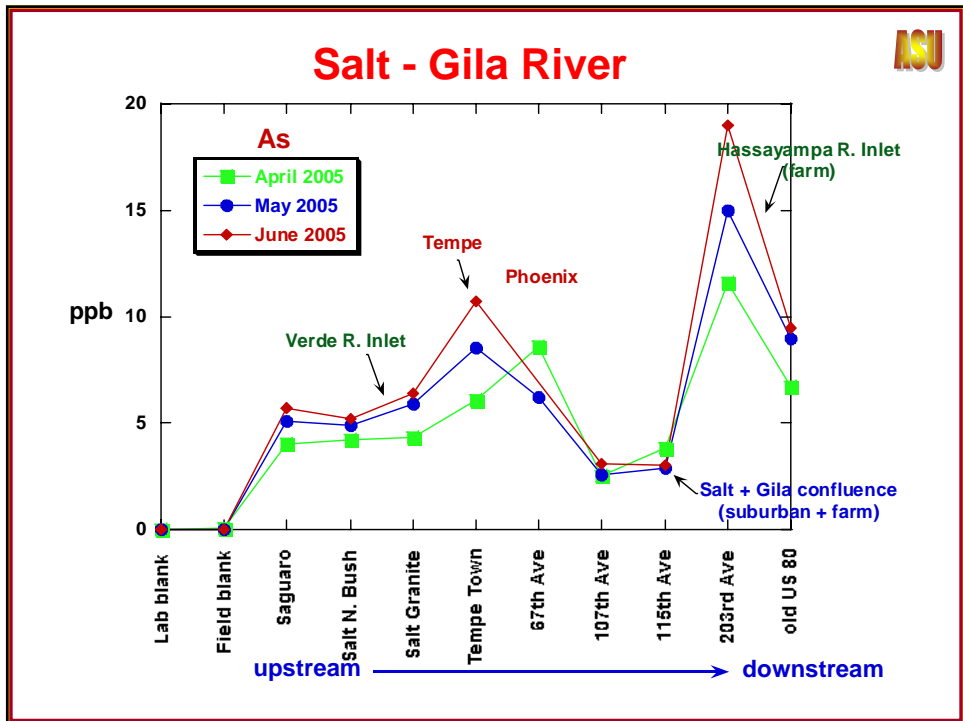
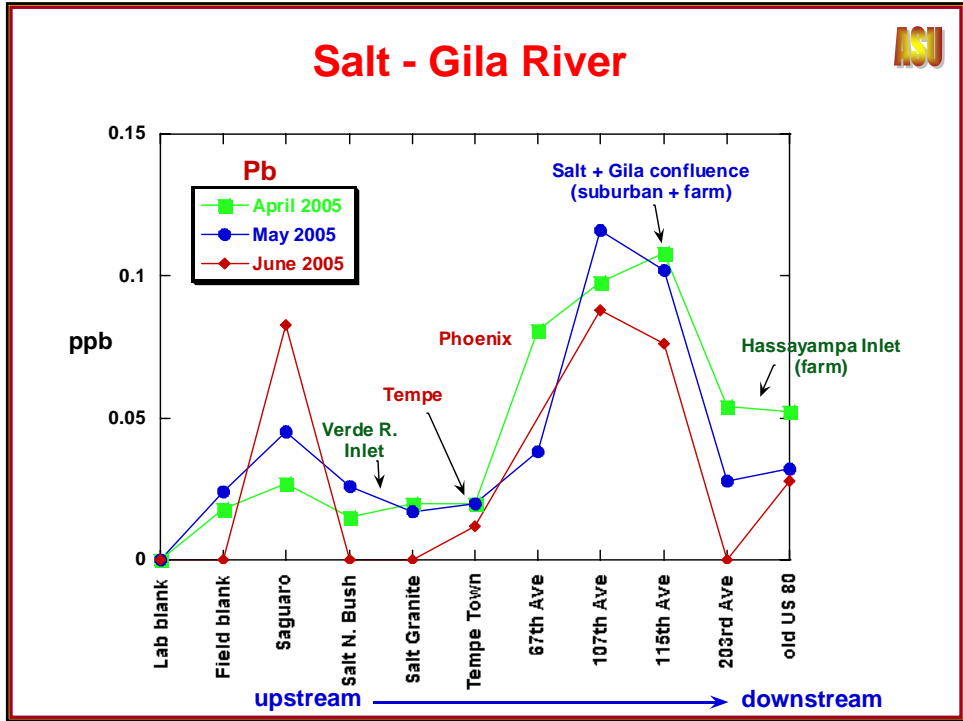
Sample Locations

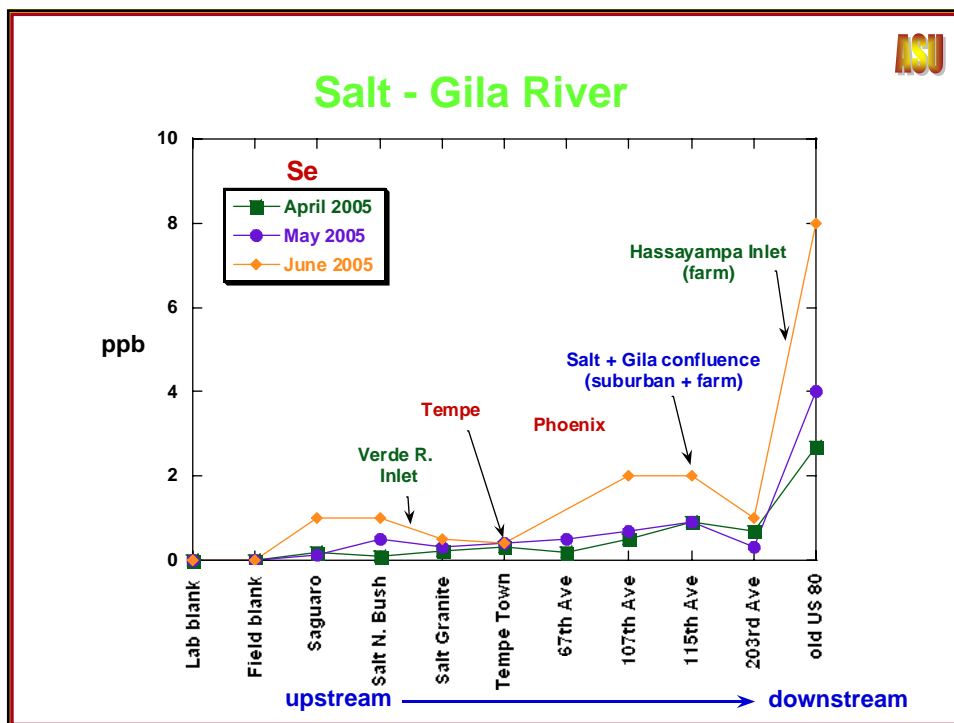
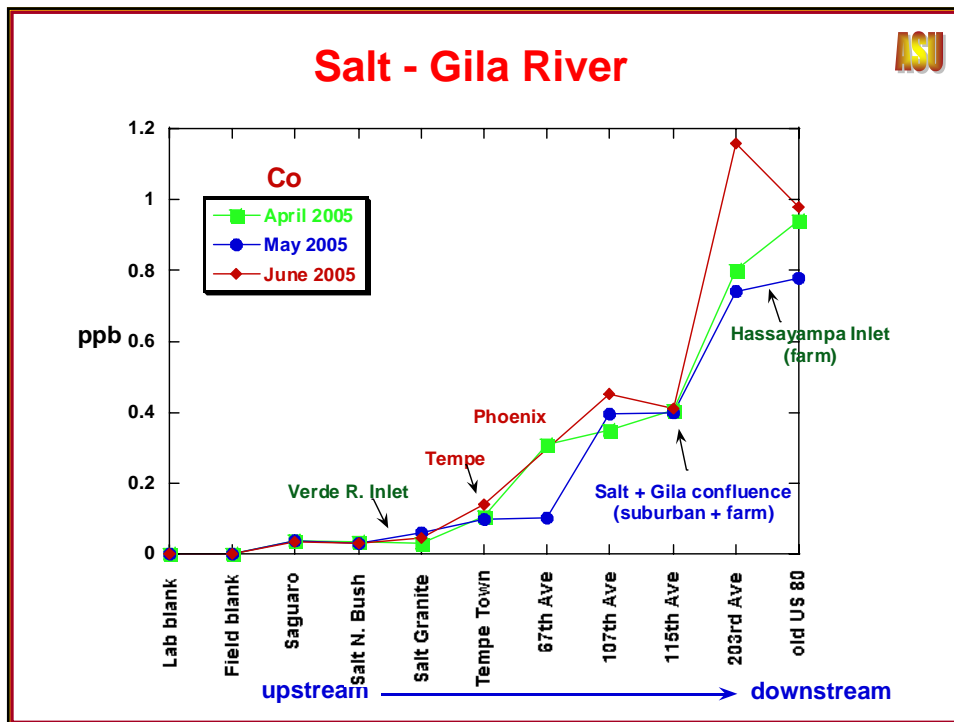


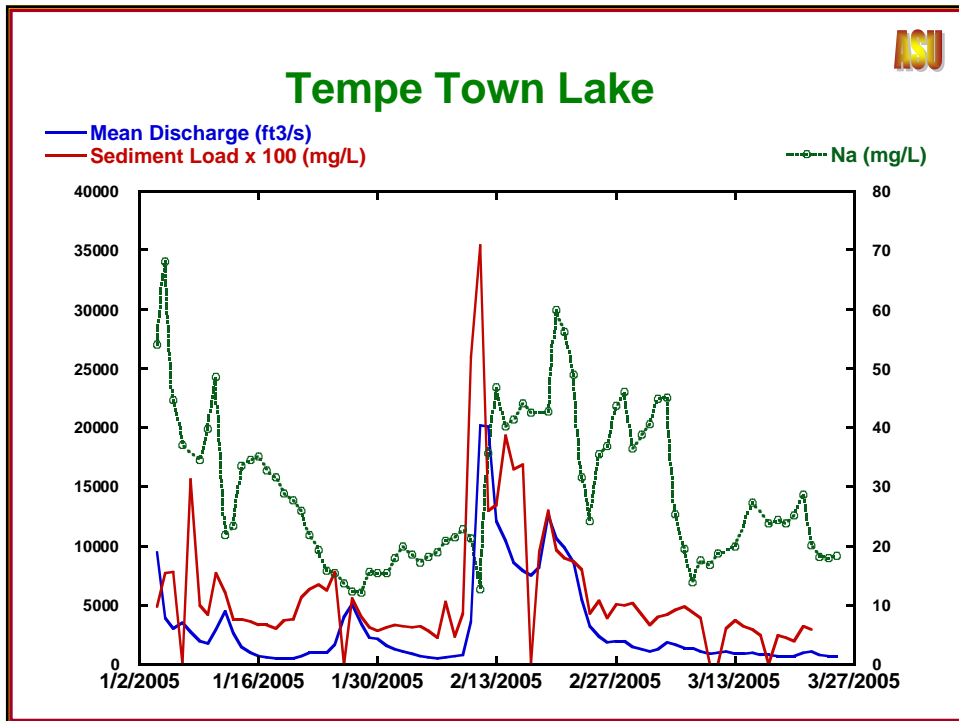
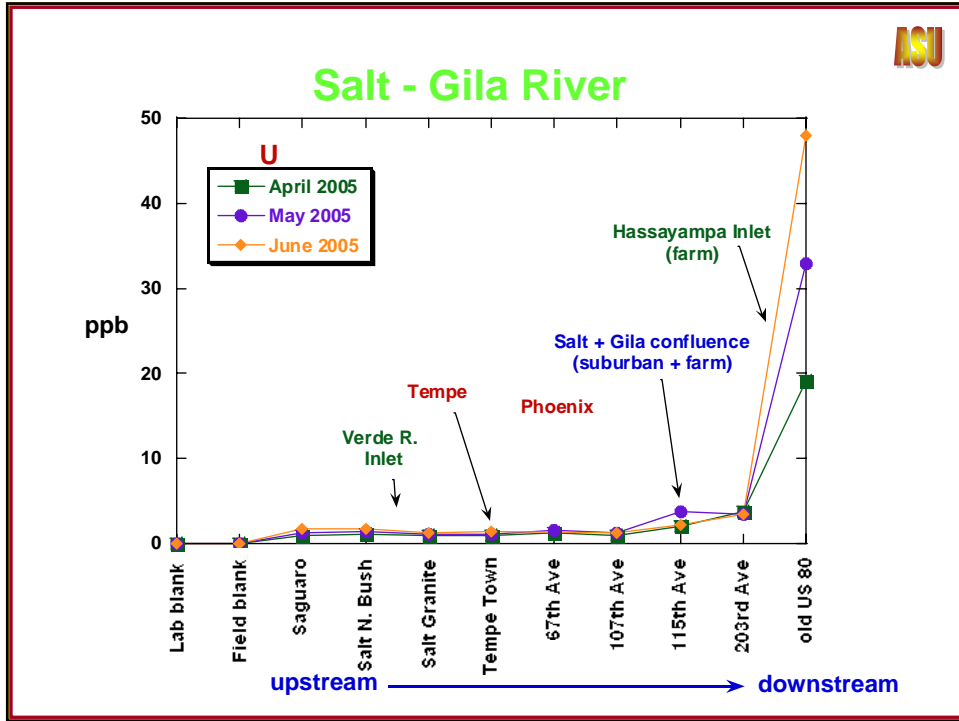
Salt River, Feb 2005

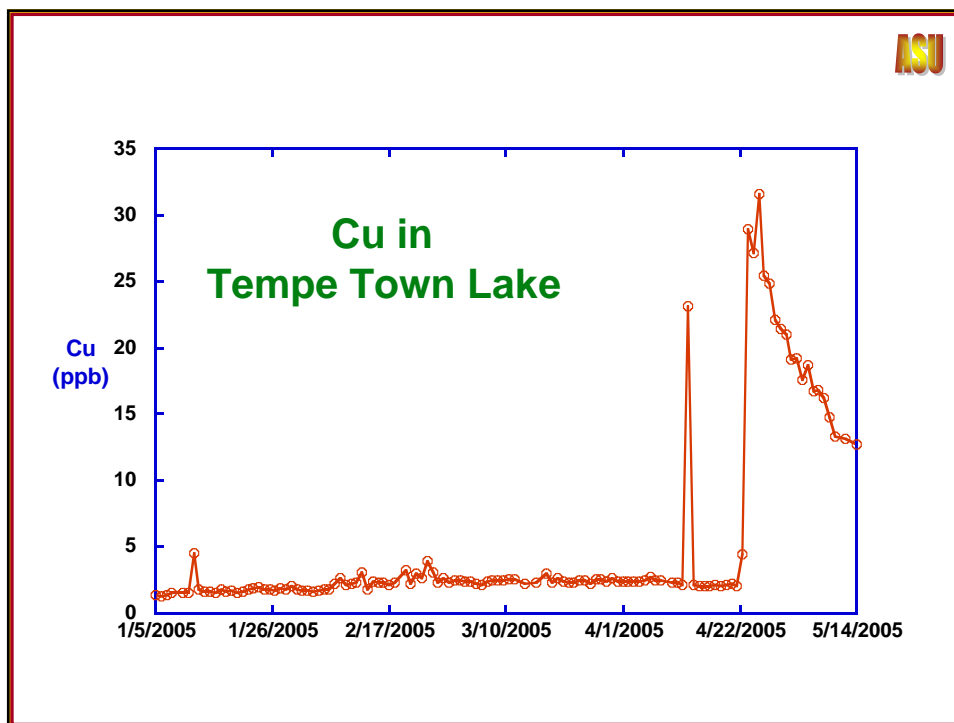
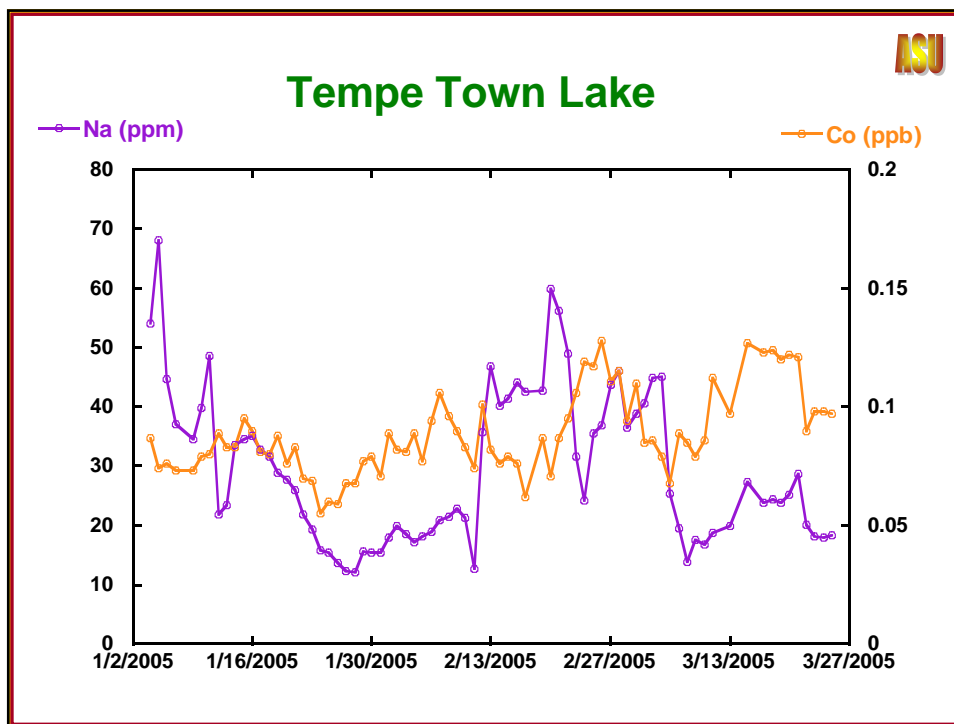


Gila River, May 2005











Acknowledgements

- **Nathan Schnebly**
- **Marisa Masles**
- **Young-II Kim**



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Fundamentals of GAC

Including T&O and DOC control

John Crittenden

**International
Institute for
Sustainability:
Consortium
Of Rapidly
Developing
Regions**



ASU Ira A. **FULTON**
school of engineering

ASU
**Issues and Technologies
Associated with Sustainable
Water Supplies**

John C. Crittenden, Ph.D., N.A.E., P.E.

Director of Consortium

Of Rapidly Developing Regions

Richard Snell Presidential Chair of Civil
and Environmental Engineering,
ASU Main Campus

Ira A. Fulton School of Engineering
Department of Civil and Environmental
Engineering

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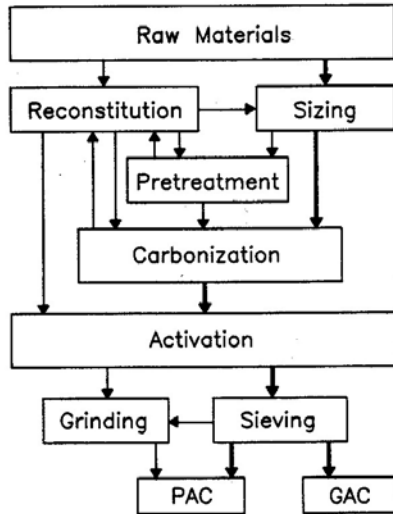
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Activated Carbon Adsorption **ASU**

Is a process whereby molecules are transferred from a fluid stream and concentrated on a solid surface by chemical (e.g., reaction, chemisorption) and physical forces (e.g. Van der Waals, physisorption).

Flow scheme to produce activated carbon

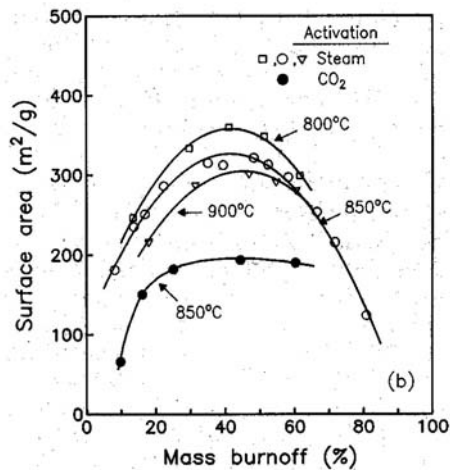
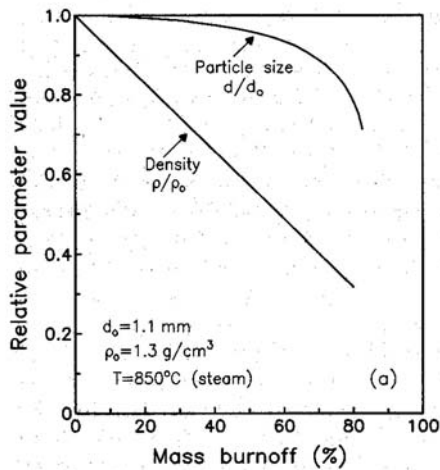
General Flow Scheme



Process Flow Description

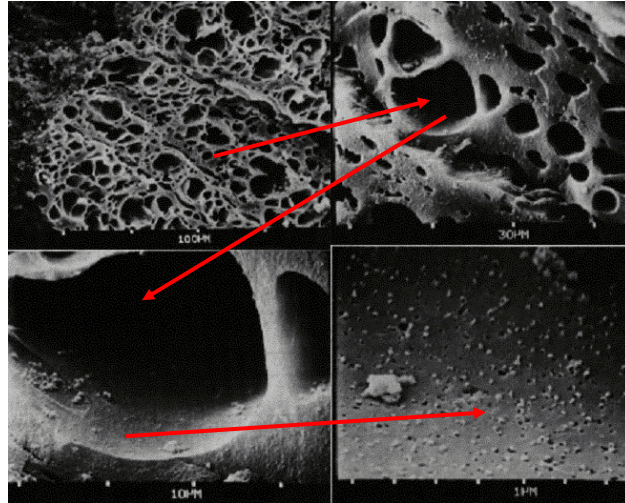
- Crushing and sizing raw materials.
- Carbonization process removes volatile components from the raw materials and realigns the carbon pore structure.
- Activation process selectively removes carbon and opens up the closed pores and increases the average size of micropores. Max. surface area per weight found at 40 -50% burnoff.
- Two Types of Activation:
 - Chemical Activation - combines carbonization and activation steps with dehydrating agents (e.g. Zinc chloride, Phosphoric acid) at 300 - 600°C to extract the cellulose.
 - Physical Activation - contacting carbonized char with gaseous agents (e.g. CO₂, air, steam) at 850 - 1100°C

Size and Pore Area as a Function of Mass Burnoff (AC become very friable with a large amount of burnoff and usually use <50%)



Activated Carbon

- Activate Carbon is carbonaceous material manufactured by a process that develops adsorptive properties.
- We can see micropores inside of macropores.

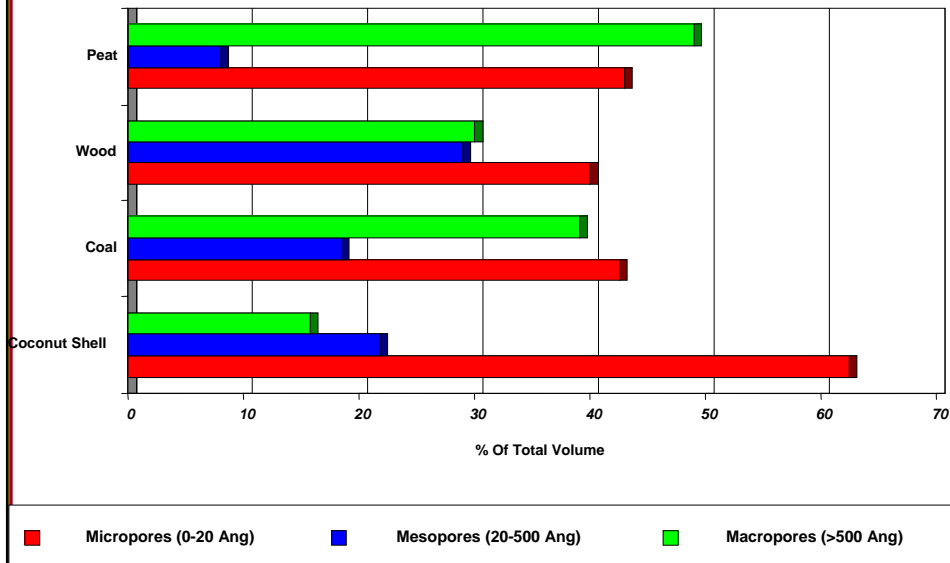


Base Material

- ◆ Coconut Shell
- ◆ Bituminous Coal
- ◆ Lignite
- ◆ Peat
- ◆ Wood
- ◆ Petroleum
- ◆ Bone Char



Pore Size Distribution Depends on ASU Several Things Including Starting Material



Activated Carbon Adsorption ASU

There are two factors that affect the adsorbability of a compound: size and solubility. Adsorbability increases with increasing size and decreasing solubility.

The factors that affect a given adsorbent's capacity for a given compound is surface chemistry and pore size. As far as surface chemistry is concerned, low ash content and the lowest possible concentration of oxygen containing functional groups will improve adsorption. Starting material affects this. Obviously, the pore size have to be appropriate. TOC requires macropore and mesopores. Some compounds that have very poor adsorbability (e.g., MTBE) can be removed with AC that has more micropores.

Experiments and or models need to be used to determine the effectiveness of AC.

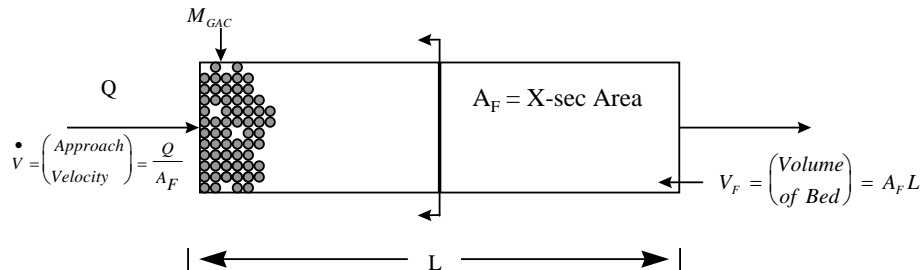
Type of Adsorption reactors

- (1) **GAC Gravity Feed Filters**
- (2) **GAC Sand Replacement Filters**
- (3) **GAC Pressure Filters**
- (4) **Biologically Active GAC Adsorbers**
- (5) **Powder activated carbon (PAC) contactors**

Activated Carbon Adsorption

Parameter	Granular activated carbon (GAC)	Powdered activated carbon (PAC)
Principal uses	Control of toxic organic compounds that are present in groundwater Barrier to occasional spikes of toxic organics in surface waters and control of taste and odor compounds Control of disinfection by- product precursors or DOC. Typical Operating Conditions: 10 to 30 min of contact time, Size: .7 to 1.3 mm.	Seasonal control of taste and odor compounds and strongly adsorbed pesticides and herbicides at low concentration (<10 ug/L). Typical Dosages 3 – 15 mg/L. Size: 10 to 70 microns.
Advantages	Easily regenerated Lower carbon usage rate per volume of water treated as compared to PAC	Easily added to existing coagulation facilities for occasional control of organics
Disadvantages	Need contactors and yard piping to distribute flow and replace exhausted carbon Previously adsorbed compounds can desorb and in some cases appear in the effluent at concentrations higher than present in the influent	Hard to regenerate and impractical to recover from sludge from coagulation facilities Much higher carbon usage rate per volume of water treated as compared to GAC

Important Variables in Fixed Bed Adsorption



Empty Bed Contact Time $t_{EBCT} = \frac{V_F}{Q} = \frac{A_F L}{\dot{V} A_F} = \frac{L}{\dot{V}}$ (6-1)

GAC Packed Bed Alternatives

GAC Sand Replacement Reactor: Replacement of sand in a filtration operation. This gives only 3-7 minutes of EBCT but the GAC seems to last for many years for Taste and Odor compounds probably because of biological degradation.

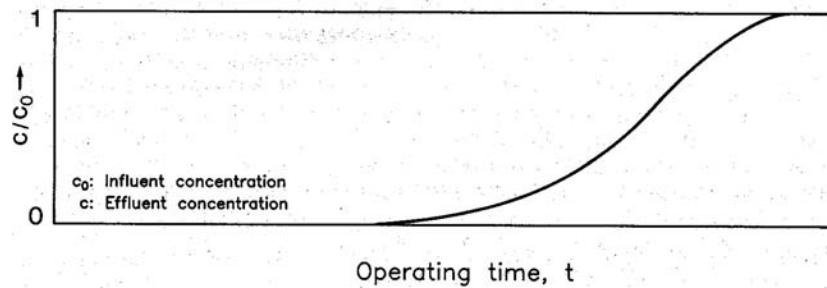
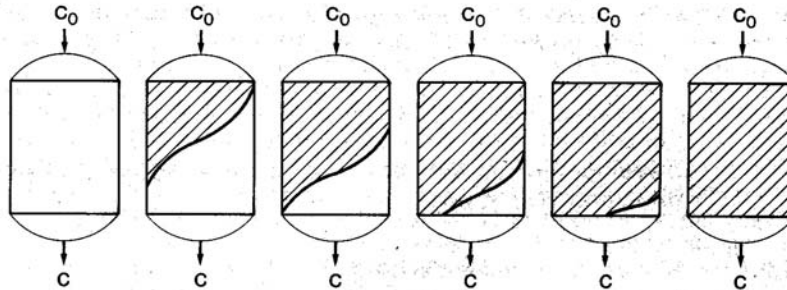
Removal of Hazardous Organic Compounds: Provide longer EBCT (10-20 minutes). The greatest amount of water treated/ mass of GAC is found for EBCTs around 10-20 minutes. Backwashing is to be avoided.

DOC/TOC removal for disinfection by product removal: The greatest amount of water treated/ mass of GAC is found for EBCT 15-45 minutes. Backwashing is to be does not seem to matter and is required.

In water treatment, often Biot is high therefore hydraulic loading does not matter for a given EBCT. (External mass transfer does not matter.) **Typical hydraulic loading is Typical filter velocities range from 5 - 15 m/h (2 - 7 gpm/ft²).**

Breakthrough Characteristics of a Fixed Bed Adsorption (Vermeulen, 1958)

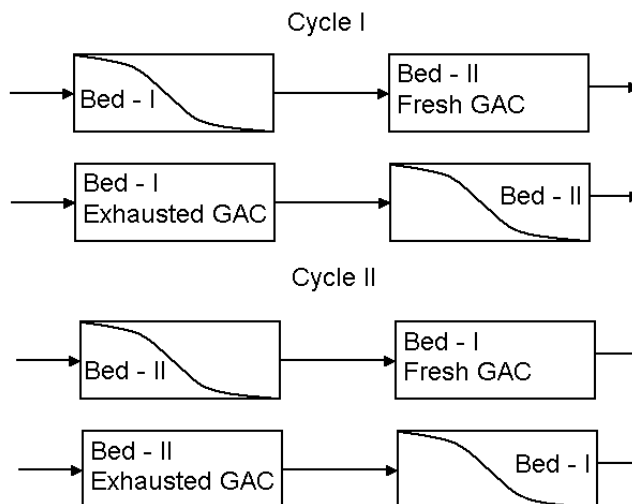
ASU



GAC Column Operation

ASU

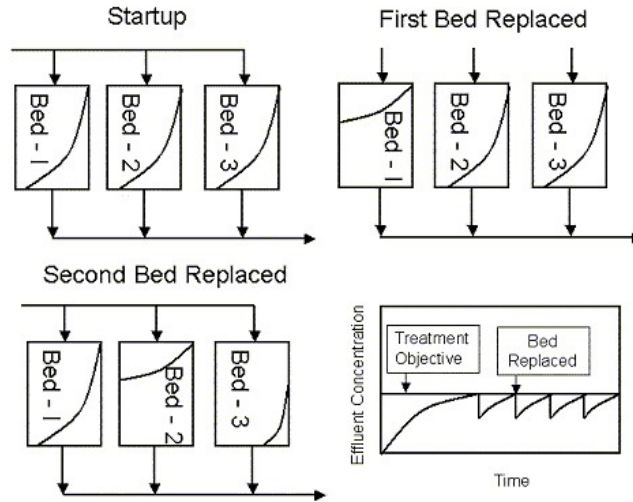
A beds in-series operation will utilize more column capacity than a single bed operation specially for less stringent treatment objectives (e.g. $C/C_{TO} < 0.05$). Increase of 20 to 50% more water treated/mass GAC.



GAC Column Operation



A beds in-parallel operation will utilize more column capacity than a single bed operation specially for less stringent treatment objectives (e.g. $C/C_{TO} > 0.3$). Increase of 50 to 150% more water treated/mass GAC depending on Treatment Objective and increases as the number of contactors in parallel is increased.



Methods for estimating full scale GAC performance



Method	Reliability	Advantages	Disadvantages
Pilot studies	Excellent	1. Can predict full scale GAC performance very accurately.	1. Can take a very long time to obtain results. 2. Expensive and must be conducted onsite.
RSSCTs	Good	1. Can predicts full scale GAC performance accurately. 2. Small volume of water is required for the test, which can be transported to a central laboratory for evaluation. 3. Can be conducted in the fraction of the time and cost that is required to conduct pilot studies.	1. Cannot predict GAC performance for different concentrations. 2. Biological degradation that may prolong GAC bed life is not considered.
Models	Fair	1. Once calibrated, models can be used to predict impact of EBCT and changes in influent concentration. 2. Can predict breakthrough of SOCs with 20 to 50 percent error.	1. Cannot predict TOC breakthrough and must be used in conjunction with pilot or RSSCT data. 2. Accurate prediction of SOC removal requires calibration with pilot or RSSCT data.

Design Strategies for TOC Removal



Consider the case study performed by Metropolitan Water District of Southern California to remove trihalomethane formation potential from their drinking water. In this case, the influent concentration is about 2.5 mg/L as TOC and the treatment objective is 1.0 mg/L as TOC which corresponds to 50 ug/L SDS THMFP.

Six participating utilities: MWD, Cincinnati, Jefferson Parish, Philadelphia, Atlanta.

Design Strategies for TOC Removal

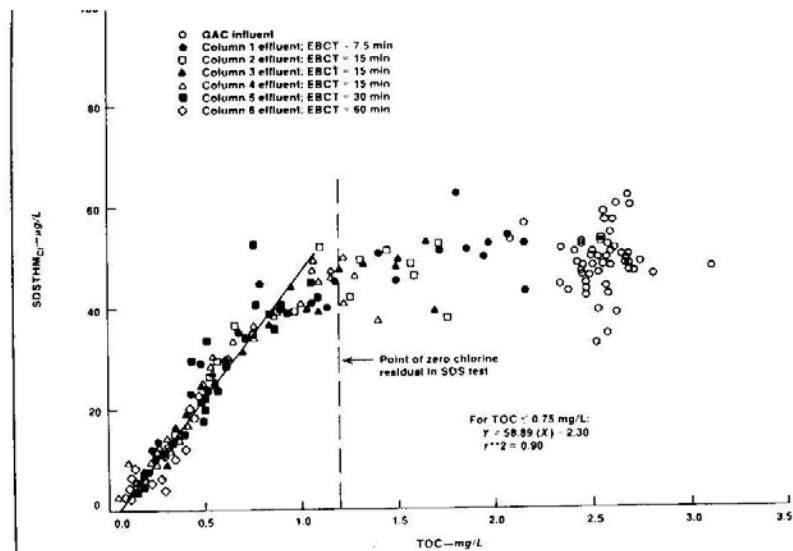
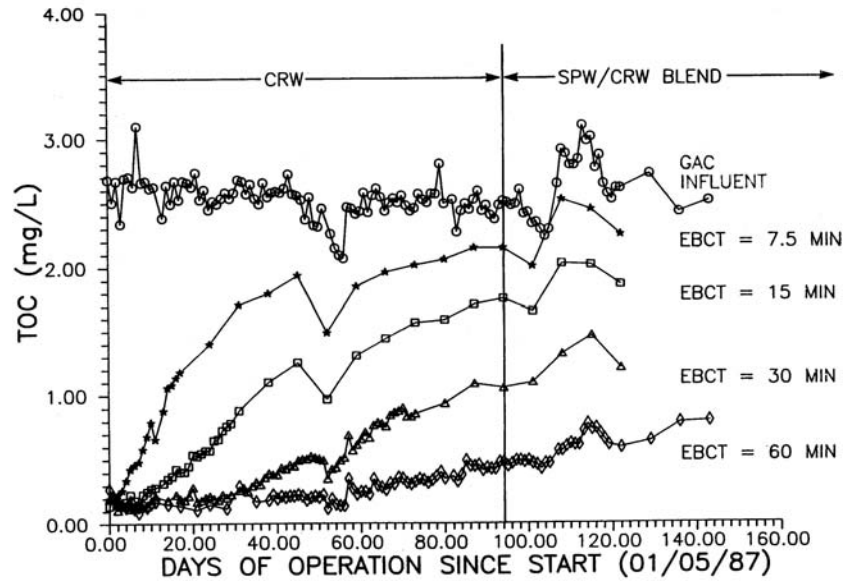
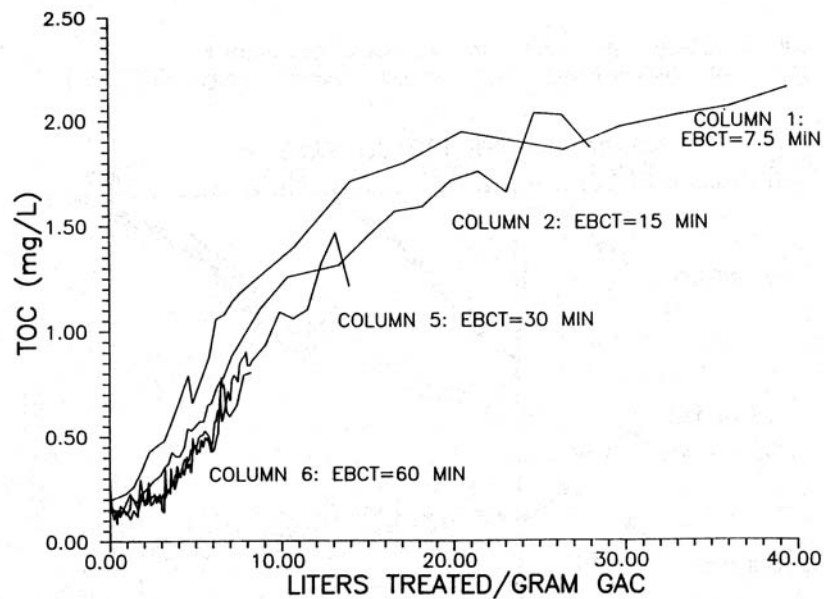


Figure 10. Linear relationship between $SDSTHM_{Cl}$ and TOC (100 percent CRW)

Design Strategies for TOC Removal – Notice the Convex downward Breakthrough Curve



Design Strategies for TOC Removal



Design Strategies for TOC Removal –
Treatment Objective = 1 mg/L = 50 ug/L of THM

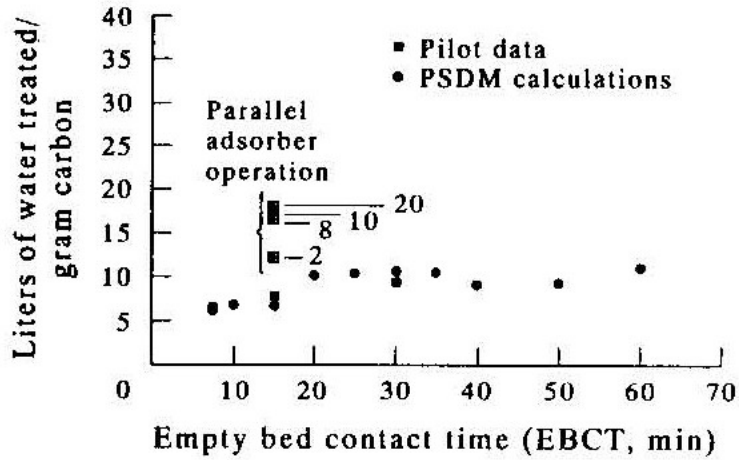
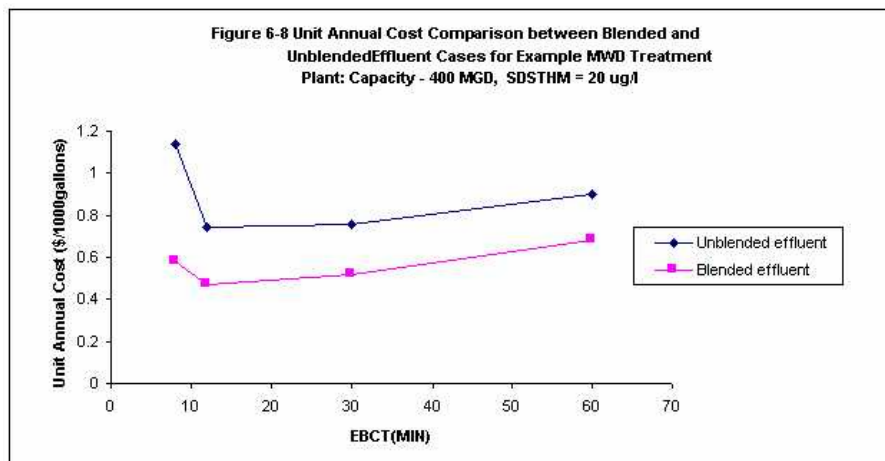


Fig. 9. Liters of water treated per gram of GAC for a DOC treatment objective of 1 mg/l as a function of EBCT for both the raw pilot data and PDM calculations.

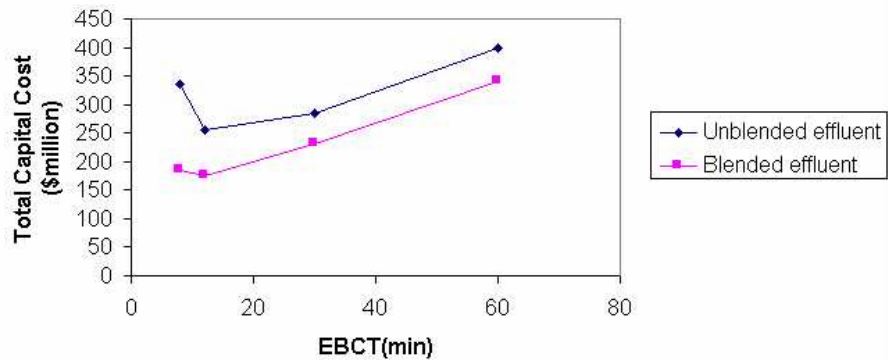
Design Strategies for TOC Removal



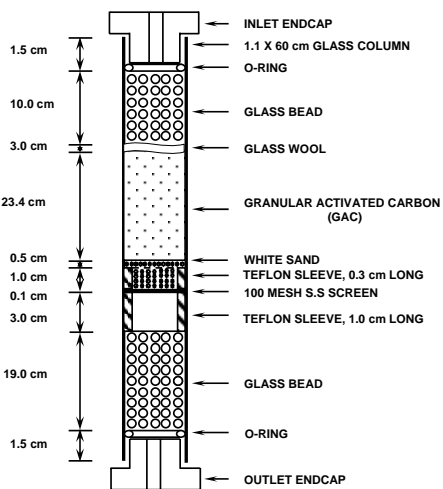
Design Strategies for TOC Removal



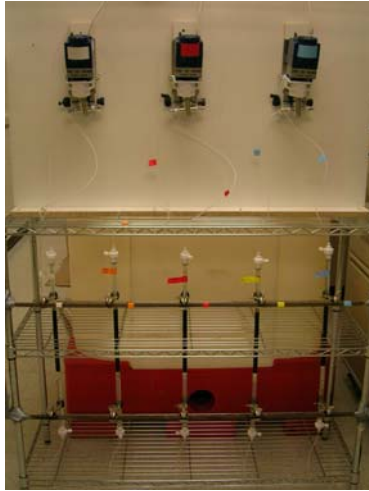
Figure 6-9 Total Capital Cost Comparison between Blended and Unblended Effluent Cases for Example MWD Treatment Plant: Capacity - 400 MGD, SDSTHM = 20 ug/l



Rapid Small Scale Column Tests (RSSCTs) can be used to assess GAC performance



Photograph of RSSCT setup with large 250-gallon feed tank behind columns



Rapid Small Scale Column Tests (RSSCTs) can be used to assess GAC performance

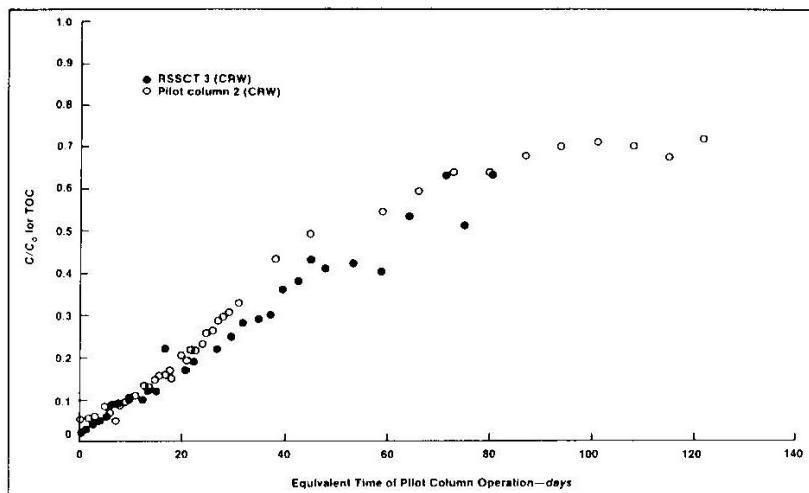


Figure 12. Comparison of TOC breakthrough profile from RSSCT with that from pilot GAC column (setup of RSSCT assumed intraparticle diffusivity to be a linear function of GAC particle size)

Rapid Small Scale Column Tests (RSSCTs) can be used to assess GAC performance

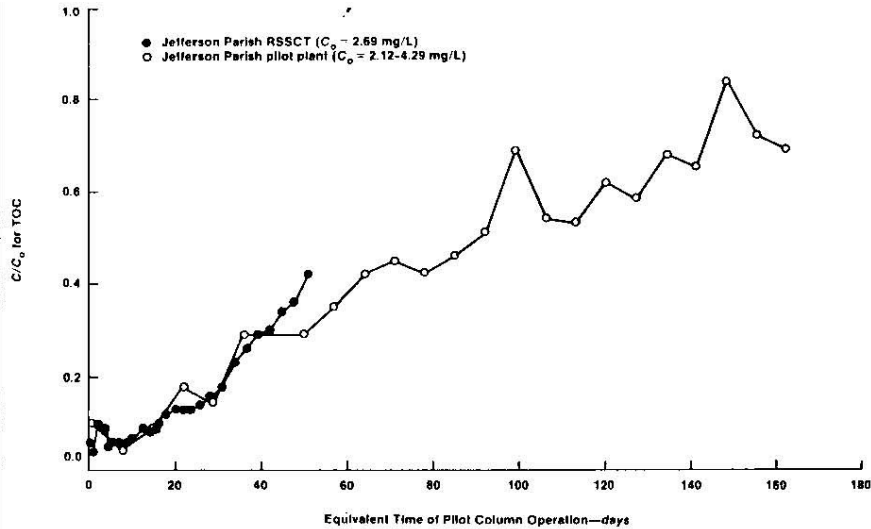


Figure 14. TOC breakthrough profiles for RSSCT and pilot plant for Jefferson Parish (RSSCT EBCT—15 min; pilot EBCT—20 min; pilot run 3)

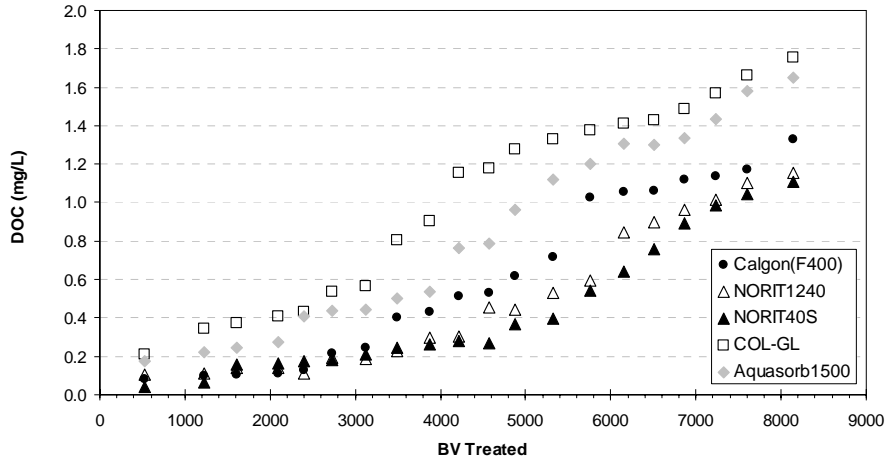


City of Scottsdale Testing by ASU

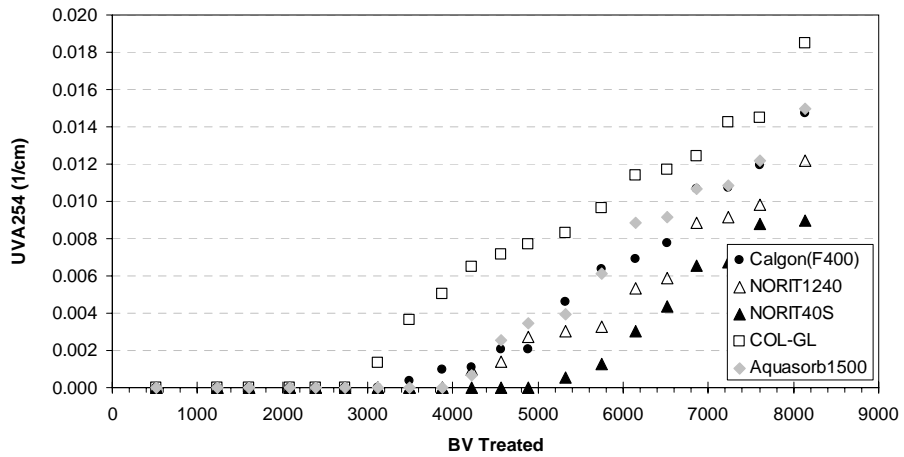
Design & operating parameters for full-scale contactor and RSSCT

Design Parameters	Full-scale	RSSCT
Particle Radius (cm)	0.0513 (12 X 40)	0.0049 (140x170)
EBCT (minutes)	20	1.91
Loading Rate (gpm/ft ²) [m/h]	4.3 (12)	3.0 (7.35)
GAC Contactor Length (ft) Width (ft) Surface Area (ft ²)	25 50 1250	1.1
RSSCT Column Diameter (cm)		
Bed Depth	10 ft (256 cm)	23.4 cm

TOC and UVA breakthrough curves from RSSCTs



TOC and UVA breakthrough curves from RSSCTs



Design Strategies for a Single Compound



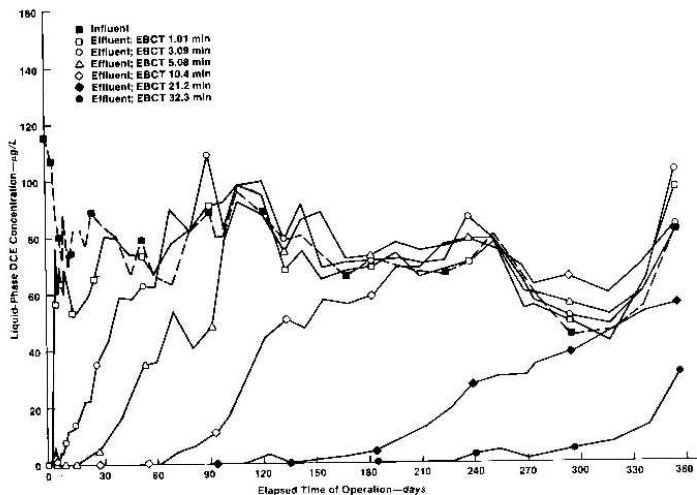
Consider a pilot plant study performed by Hand et al. (1989). The following table summarizes the organic compounds to the column and their average influent concentrations from well no. 4 (Hand et al., 1989).

Volatile Organic Compound	Number of Data	Average Influent Concentration ($\mu\text{g/L}$)	Standard Deviation ($\mu\text{g/L}$)
Vinyl Chloride	41	8.2	2.4
1,1,1-Trichloroethane	44	0.9	0.5
Cis-1,2-Dichloroethene	44	70.9	19.0
Trichloroethene	44	47.9	22.1
Tetrachloroethene	44	37.6	17.6
Toluene	36	19.3	11.7
Ethyl Benzene	35	4.5	.9
P-Xylene	37	5.2	1.7
O and P Xylene	38	9.3	3.0

Design Strategies for a Single Compound



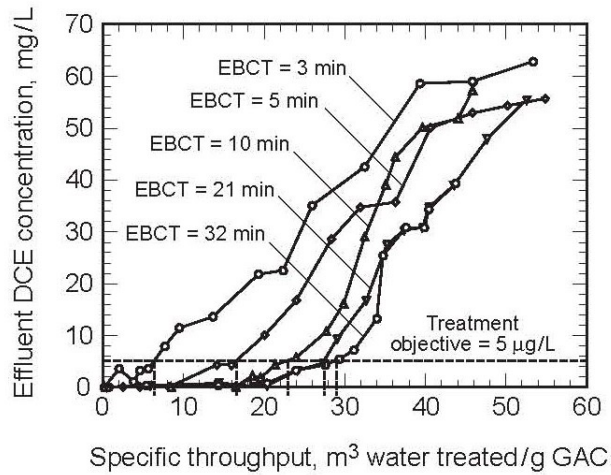
Consider the following effluent profiles for cis-1,2-dichloroethene (DCE) taken from a pilot plant study in Wausau, WI (Hand et al., 1989). Notice the shape to the Breakthrough Curve.



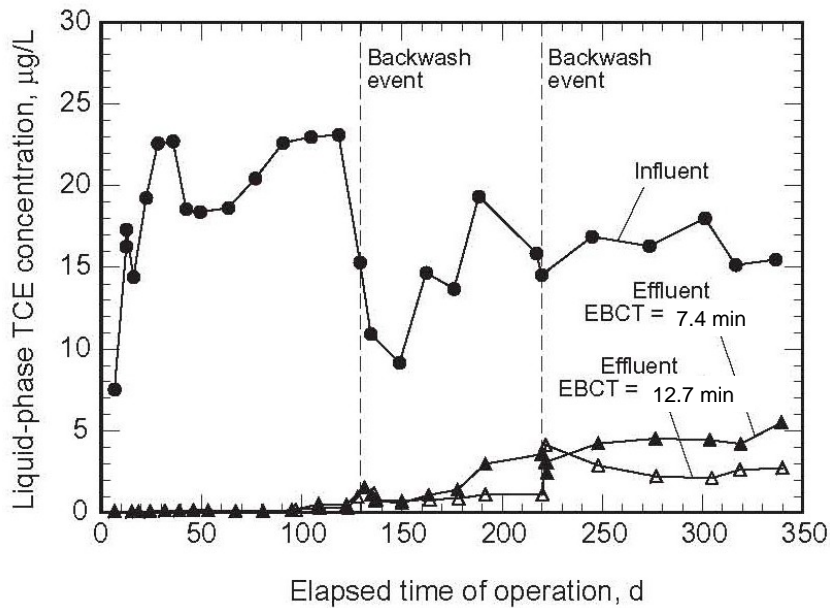
Design Strategies for a Single Compound



Consider the following effluent profiles for **cis-1,2-dichloroethene (DCE)** taken from a pilot plant study in Wausau, WI (Hand et al., 1989).



Impact of Backwashing for SOC Removal



Model Predicted Removal



	k	unit	n		
MIB	17.6	ng/mg(L/ng) ^{1/n}	0.34	184.2946	ug/g(L/ug) ^{1/n}
Geosmin	34.5	ng/mg(L/ng) ^{1/n}	0.35	387.0964	ug/g(L/ug) ^{1/n}

Source: Graham, Wat. Res., vol 34, 8, pp2291-2300, 2000

The screenshot shows the AdDesignS software interface with the following parameters:

- Water Properties:** Pressure = 1.00000 atm, Temperature = 15.0 C.
- Component Properties:** MIB, Geosmin.
- Simulation Parameters for PSDM Only:** Total Run Time = 50.0 year, First Point Displayed = 13.2 d, Time Step = 0.200 year, Number of Axial Elements = 1, Axial Direction = 8, Radial Direction = 3.
- Fixed Bed Properties:** Bed Length = 2.77 m, Bed Diameter = 3.05 m, Bed Mass = 9072 kg, Flowrate = 0.0225 m³/s, EBCT = 15.0 min, Bed Density = 0.4485 g/mL, Bed Porosity = 0.441, Superficial Velocity = 11.034 m/hr, Interstitial Velocity = 25.130 m/hr.
- Adsorbent Properties:** Name = Calgon F400 (12x40), Apparent Density = 0.803 g/mL, Particle Radius = 5.13E-04 m, Porosity = 0.641, Particle Shape Factor = 1.00.

EBCT = 7.5min



Results for the PSDM (No Reactions Present)

The screenshot shows the results window for the PSDM simulation. It includes a table of results and a graph of C/Co vs Time(days).

	Time (days)	BVT(m ³ /m ³)	VTM(m ³ /kg)	C (mg/L)
5% of influent conc.	N/A	N/A	N/A	N/A
50% of influent conc.	N/A	N/A	N/A	N/A
95% of influent conc.	N/A	N/A	N/A	N/A
Treatment Objective	N/A	N/A	N/A	N/A

The graph shows C/Co on the y-axis (0.0 to 0.9) and Time(days) on the x-axis (0 to 1500). Two curves are shown: MIB (blue) and Geosmin (green). Both curves show a sharp increase in C/Co starting around 500 days, reaching approximately 0.85 by 1000 days. The MIB curve is slightly higher than the Geosmin curve.

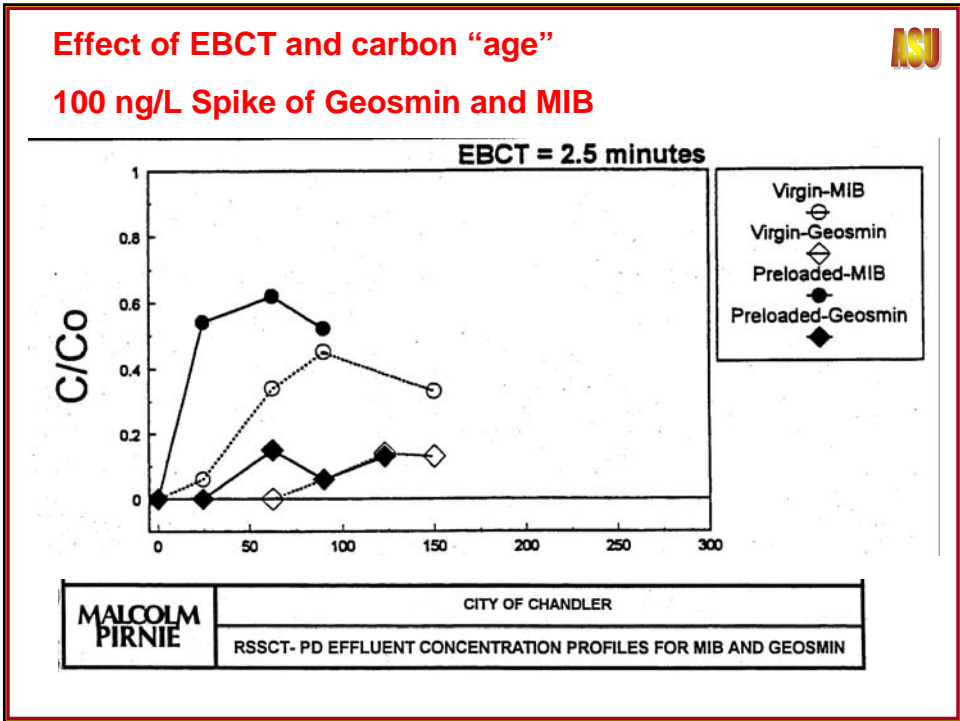
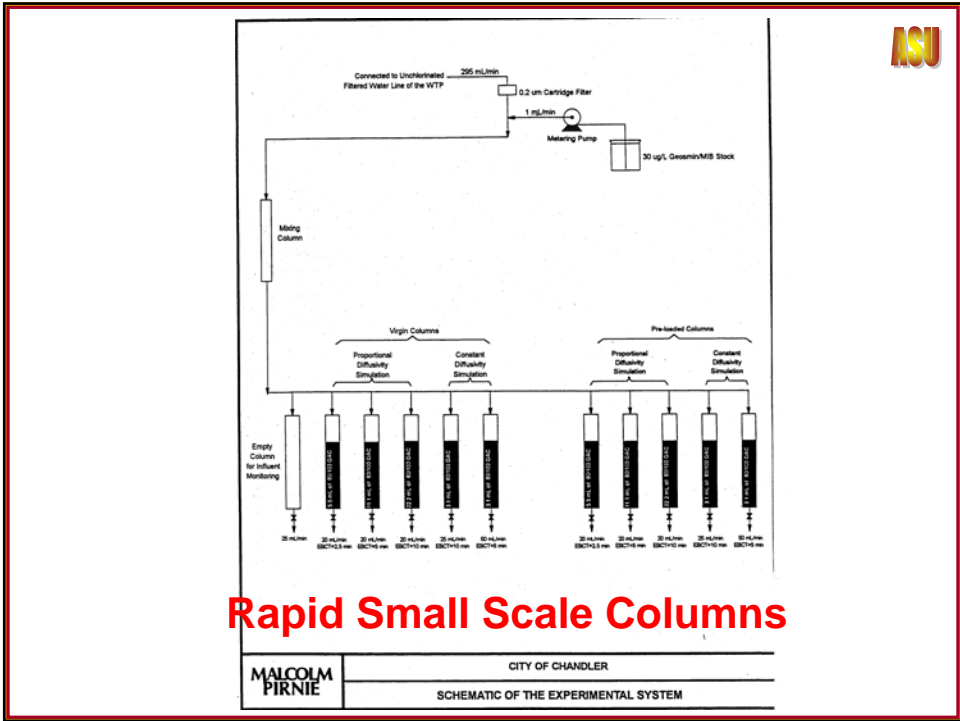
GAC Caps for MIB Control

- GAC Filter caps
- Replace anthracite layer in dual media filters; sand layer remains
- Provides short-term adsorption for DOC (DBP precursors) and MIB / Geosmin
- Provides sustainable removal via biodegradation
- Would have to delay point of chlorination to have biologically active GAC
- MPI pilot study; Chandler WTP experience



Chandler WTP

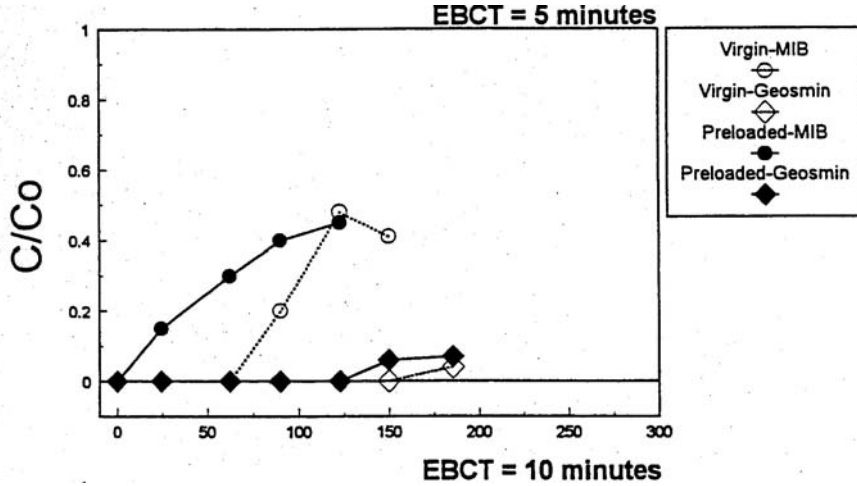
- **Full-Scale Filter Design:**
 - Filter caps designed to act as a barrier against synthetic organic compounds and T&O compounds
 - 20 inches GAC (8x30 US mesh size, Elf Atochem)
 - 10 inches of sand support
 - 12 inches of graded gravel
 - GAC replaced every 3 years @ HLR = 4 gpm/sf Purpose of the study:
 - It was planned to increase the capacity the WTP by increasing the rate of filtration from 4 to 6 gpm/sf
 - Evaluate the effect of a higher filtration rate on the removal of T&O compounds
 - RSSCT Tests
 - Modeling and cost estimates



Effect of EBCT and carbon "age"



100 ng/L Spike of Geosmin and MIB

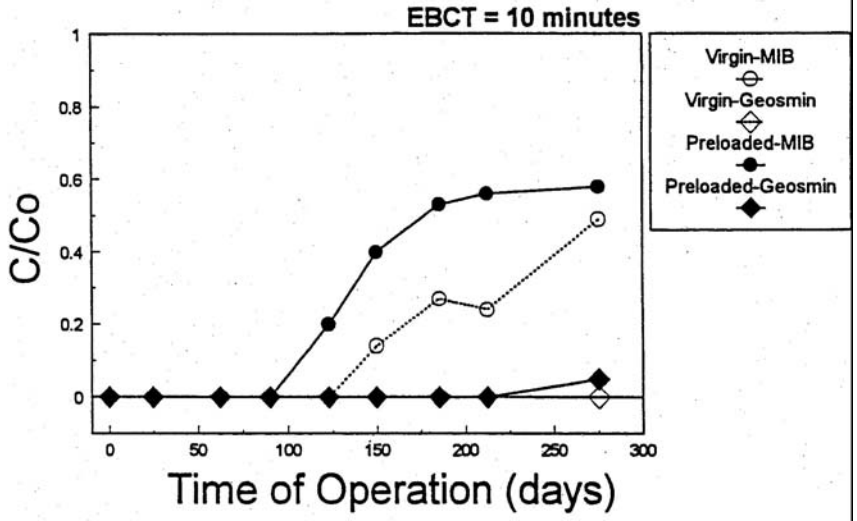


MALCOLM PIRNIE CITY OF CHANDLER
RSSCT- PD EFFLUENT CONCENTRATION PROFILES FOR MIB AND GEOSMIN

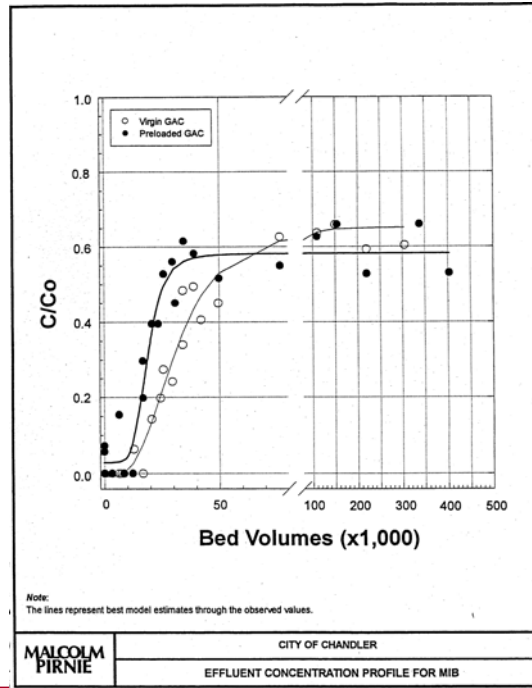
Effect of EBCT and carbon "age"



100 ng/L Spike of Geosmin and MIB

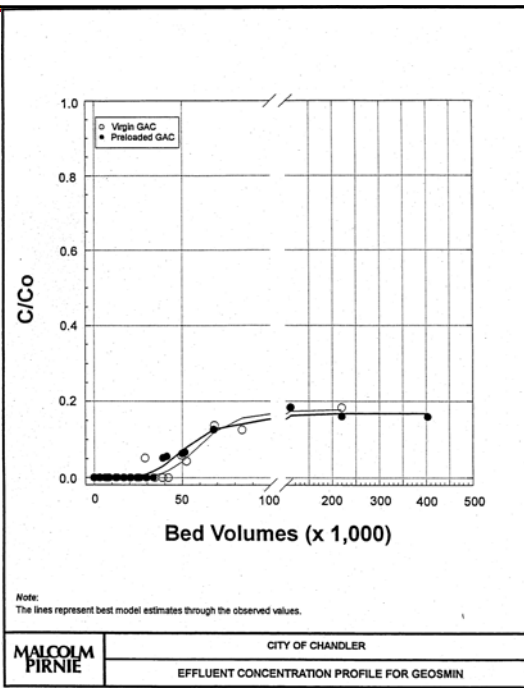


MALCOLM PIRNIE CITY OF CHANDLER
RSSCT- PD EFFLUENT CONCENTRATION PROFILES FOR MIB AND GEOSMIN



MIB

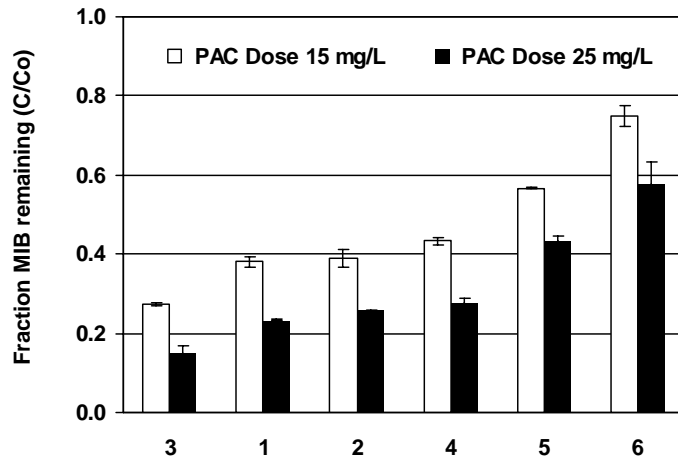
- 40% sustainable removal
- 1000 bed volumes ~ 1 day



Geosmin

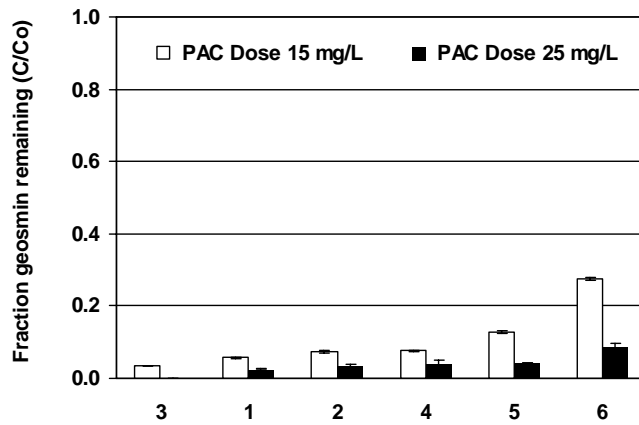
- 80% sustainable removal

Fraction MIB and geosmin remaining according to Powder Activated Carbon (PAC)



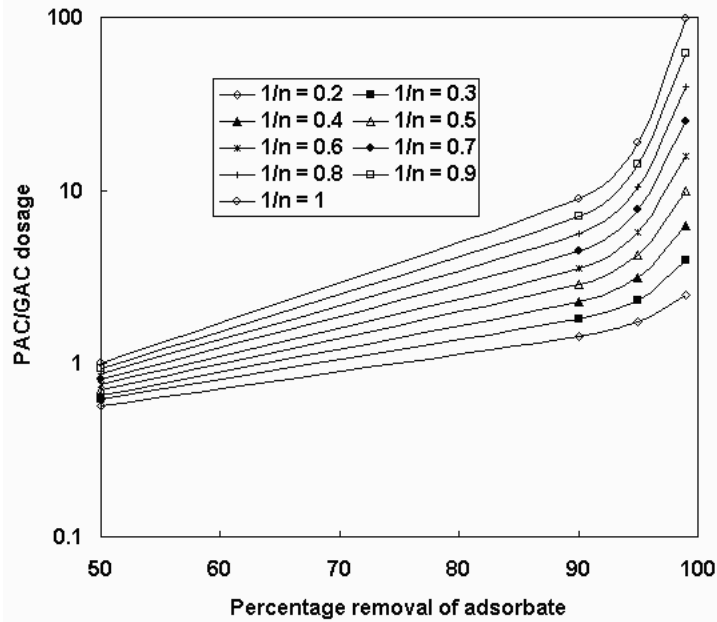
NO	source
1	wood
2	wood
3	-
4	Coal
5	Lignite
6	Bituminous

Fraction MIB and geosmin remaining according to Powder Activated Carbon (PAC)



NO	source
1	wood
2	wood
3	-
4	Coal
5	Lignite
6	Bituminous

Usage Rates GAC versus PAC



Water Treatment Book - Theory Reduced to Practice Montgomery-Watson-Harza invested \$ 1 million, 1948 Pages



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2	Physical and Chemical Quality	17 Reverse Osmosis	1429
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5	Fundamentals of Chemical Reactions	20 Residuals Management	1641
6	Reactor Analysis	21 Internal Corrosion of Water Conduits	1709
7	Introduction to Separation Processes and Mass Transfer	22 Synthesis of Treatment Trains: Case Studies From Bench to Full Scale	1819
8	Chemical Oxidation and Reduction		

Activated Carbon Summary



- There are two factors that affect the adsorbability of a compound: size and solubility. Adsorbability increases with increasing size and decreasing solubility.
- The factors that affect a given adsorbent's capacity for a given compound is surface chemistry and pore size. As far as surface chemistry is concerned, low ash content and the lowest possible concentration of oxygen containing functional groups will improve adsorption. Starting material affects this. Obviously, the pore size have to be appropriate. TOC requires macropore and mesopores. Some compounds that have very poor adsorbability (e.g., MTBE) can be removed with AC that has more micropores.
- GAC has a much lower carbon usage rate per volume of water treated as compared to PAC; however, contactors and yard piping is Needed to distribute flow and replace exhausted carbon
- PAC can easily be added to existing coagulation facilities for occasional control of organics such as MIB and Geosmin but very high dosages of PAC are required to control MIB.

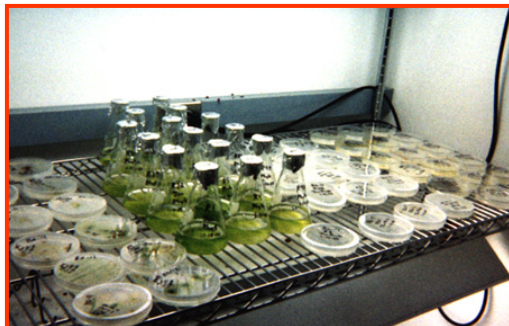
Activated Carbon Summary

ASU

- GAC:
 - ◆ TOC and DBP precursors can only be controlled using GAC
 - ◆ In terms of reliability and time requirements to predict GAC effectiveness, this is the order: Pilot, RSSCTs, and mathematical models.
 - ◆ Beds in parallel can reduce the GAC usage rate for treatment objectives greater than ~30%
 - ◆ Beds in series can reduce the GAC usage rate for treatment objectives less than ~5%
 - ◆ Backwashing is detrimental to GAC treatment of hazardous organics and not important for DOC
 - ◆ GAC can be effective as a sand filter replacement for biological and long term removal of taste and odor compounds

Recent Advances For Early Warning Algae Systems

ASU





DNA-Based Sensor for T&O- and Toxin-Producing Algae

Qiang Hu¹, Milton Sommerfeld¹, and Paul Westerhoff²

¹School of Life Sciences

**²Dept. of Civil and Environmental Engineering
Arizona State University**

**Project supported by:
Salt River Project
ASU Water Quality Center**

September 2, 2005



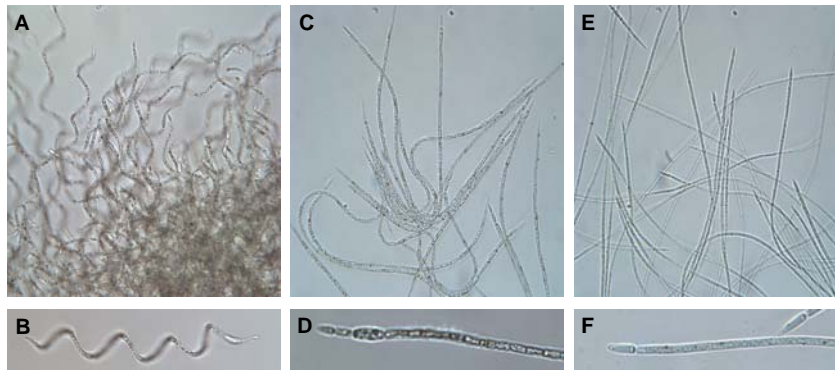
Goal of Research

To develop a PCR-based DNA fingerprinting method for rapid, sensitive, and reliable detection of potential toxin-producing cyanobacteria, and integrate the method into current water quality monitoring and management practices

Specific Objectives

- Design PCR primers specific for *Cylindrospermopsis* and other potential toxin-producing cyanobacterial species/strains
- Develop an optimized real-time PCR protocol for quantitative detection of *Cylindrospermopsis* and other toxin-producing cyanobacteria
- Use the Arizona Canal and Saguaro Lake as field experimental systems to validate the PCR and real-time PCR methods developed

Isolation of *Cylindrospermopsis* sp. from the Arizona Canal and Saguaro Lake



Light photomicrographs of *Cylindrospermopsis raciborskii* isolates

- A, B = trichomes (coiled form)
C, D = trichomes (straight form)
E, F = trichome cells (without gas vesicles)

Cylindrospermopsis raciborskii Isolates

<table border="0" style="width: 100%;"> <tr><td style="background-color: #fce4d6;">AZ-82</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-83</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-84</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-85</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-86</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-87</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-88</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-89</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-11</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-33</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr><td style="background-color: #fce4d6;">AZ-60</td><td style="background-color: #fce4d6;"><i>Cylindrospermopsis raciborskii</i> (straight)</td></tr> <tr style="background-color: #bbdefb;"><td>AZ-14</td><td><i>Cylindrospermopsis raciborskii</i> (straight, no gas vesicle)</td></tr> </table>	AZ-82	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-83	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-84	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-85	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-86	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-87	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-88	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-89	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-11	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-33	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-60	<i>Cylindrospermopsis raciborskii</i> (straight)	AZ-14	<i>Cylindrospermopsis raciborskii</i> (straight, no gas vesicle)	<table border="0" style="width: 100%;"> <tr><td style="background-color: #e1f5fe;">AZ-5</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-19</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-30</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-50</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-51</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-52</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-53</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-54</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-76</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-80</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> <tr><td style="background-color: #e1f5fe;">AZ-81</td><td style="background-color: #e1f5fe;"><i>Cylindrospermopsis raciborskii</i> (curved)</td></tr> </table>	AZ-5	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-19	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-30	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-50	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-51	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-52	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-53	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-54	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-76	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-80	<i>Cylindrospermopsis raciborskii</i> (curved)	AZ-81	<i>Cylindrospermopsis raciborskii</i> (curved)
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AZ-11	<i>Cylindrospermopsis raciborskii</i> (straight)																																														
AZ-33	<i>Cylindrospermopsis raciborskii</i> (straight)																																														
AZ-60	<i>Cylindrospermopsis raciborskii</i> (straight)																																														
AZ-14	<i>Cylindrospermopsis raciborskii</i> (straight, no gas vesicle)																																														
AZ-5	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-19	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-30	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-50	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-51	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-52	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-53	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-54	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-76	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-80	<i>Cylindrospermopsis raciborskii</i> (curved)																																														
AZ-81	<i>Cylindrospermopsis raciborskii</i> (curved)																																														

Alignment of 16S rRNA gene segments among *Cylindrospermopsis* and other cyanobacterial species

```

Planktothrix      ATGCAAGTCGAACGGAATCCTTCGGGATTTAGTGGCGGACGGGTGAGTAACACGTAAGAA
Anabaena 7120     ATGCAAGTCGAACG--GTCTCTTCGGAGATAGTGGCGGACGGGTGAGTAACGCGTGAGAA
Calothrix         ATGCAAGTCGAACGG--TACCTTCGGGTATAGTGGCGGACGGGTGAGTAACGCGTGAGAA
Microcystis      ATGCAAGTCGAACGGGAATCTTCGGATTCCAAGTGGCGGACGGGTGAGTAACGCGTAAGAA
Microcoleus      ATGCAAGTCGAACG-CAACCTTCGGGTGAG-TGGCGGACGGGTGAGTAACGCGTGAGAA
Anabaenopsis     ATGCAAGTCGAACG--GTCTTTTCGGAGATAGTGGCGGACGGGTGAGTAACGCGTGAGAA
Aph. gracile     CATCAAGTCGAACGGTCTTTTCGG--AGATAGTGGCGGACGGGTGAGTAACGCGTAAGAA
Aph. flos-aquae  ATGCAAGTCGAACG--GTCTTTTAGGAGACAGTGGCGGACGGGTGAGTAACGCGTAAGAA
Nostoc sp.       ATGCAAGTCGAACG--GTGTCTTCGGACACAGTGGCGGACGGGTGAGTAACGCGTGAGAA
Anabaena bergii  ATGCAAGTCGAACGGTCTTTTCGG--AGATAGTGGCGGACGGGTGAGTAACGCGTGAGAA
Nodularia sp.   ATGCAAGTCGAACGGTCTCTTCGG--AGATAGTGGCGGACGGGTGAGTAACGCGTGAGAA
CR (Straight)    ATGCAAGTCGAACGGGATGCTTAGGCATCTAGTGGCGGACGGGTGAGTAACGCGTGAGAA
CR (coiled)      ATGCAAGTCGAACGGGATGCTTAGGCATCTAGTGGCGGACGGGTGAGTAACGCGTGAGAA
CR (Florida)     ATGCAAGTCGAACGGGATGCTTAGGCATCTAGTGGCGGACGGGTGAGTAACGCGTGAGAA
.:*****      :      .:*****

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Cyl 16S-I

PCR Detection of *Cylindrospermopsis* in Saguaro Lake Samples



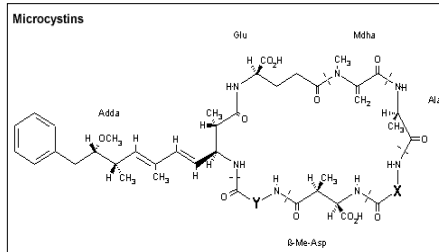
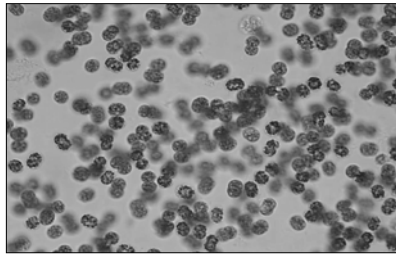
- | | |
|--|--|
| <p>1: <i>Anabaena</i> TAC426</p> <p>2: Saguaro Lake sample</p> <p>3: <i>Nodularia</i> strain 575</p> <p>4: <i>Cylindrospermopsis</i> AWT205</p> <p>5: <i>Plankothrix</i> PCC7811</p> | <p>6: <i>Microcystis</i> LE-3</p> <p>7: <i>Nostoc</i> PCC73102</p> <p>8: Saguaro Lake sample</p> <p>9: <i>Aphanizomenon</i> strain Zayi</p> <p>10: No DNA sample</p> |
|--|--|

LC/MS/MS Screening of Toxins from Isolated *Cylindrospermopsis* Strains

ID #	AZ-6	AZ-14	AZ-19	AZ-30	AZ-33
Description	Cylindro	Cylindro	Cylindro	Cylindro	Cylindro
Analyte	ppb	ppb	ppb	ppb	ppb
Cylindrospermopsin	1.2	<0.5	<0.5	<0.5	0.96
Anatoxin-a Isomer	<0.5	<0.5	<0.5	<0.5	<0.5
Microcystin RR	<0.5	<0.5	<0.5	<0.5	<0.5
Microcystin LR	<0.5	<0.5	<0.5	<0.5	<0.5
Microcystin YR	<0.5	1.5	<0.5	<0.5	<0.5
Nodularin	<0.5	<0.5	<0.5	<0.5	<0.5

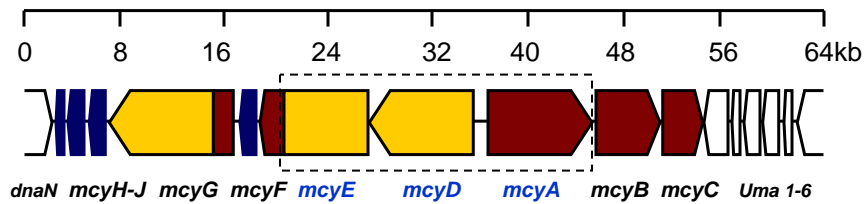
ID #	A-12	AZ-16	AZ-22	AZ-60	AZ-70
Description	Cylindro	Cylindro	Cylindro	Cylindro	Cylindro
Analyte	ppb	ppb	ppb	ppb	ppb
Cylindrospermopsin	<0.5	<0.5	~ 2.5	<50	<0.5
Anatoxin-a Isomer	<0.5	<0.5	<0.5	<0.5	<0.5
Microcystin RR	<0.5	<0.5	<2.5	<0.5	<0.5
Microcystin LR	<0.5	<0.5	<2.5	~ 50	<0.5
Microcystin YR	<0.5	1.1	<2.5	<0.5	<0.5
Nodularin	<0.5	<0.5	<2.5	<0.5	<0.5

Isolation and identification of *Microcystis* sp. from Saguaro Lake



Elisa analysis: 1.21mg *microcystin-LR*/g cell dry weight

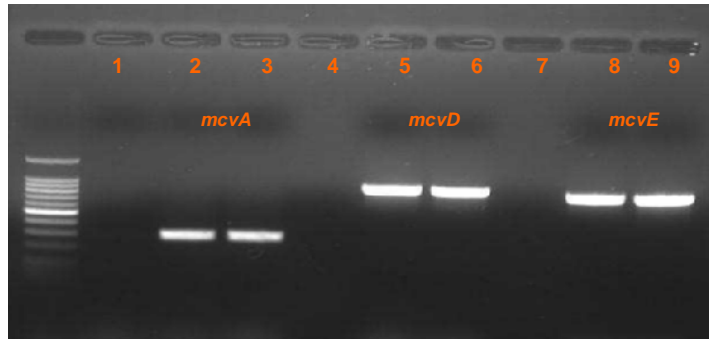
Organization of the Gene Cluster for Microcystin Biosynthesis



- Regions homologous to nonribosomal peptide synthetases
- Regions homologous to polyketide synthetases
- ORFs of putative microcystin tailoring function
- Non-microcystin synthetase ORFs

PCR Detection of *Microcystis* sp. from Saguaro Lake Samples

ASU



Lanes 1, 4, & 7: Negative controls
Lanes 2, 5, & 8: *Microcystis* sp. AZ-93 gene segments
Lanes 3, 6, & 9: *Microcystis* positive controls

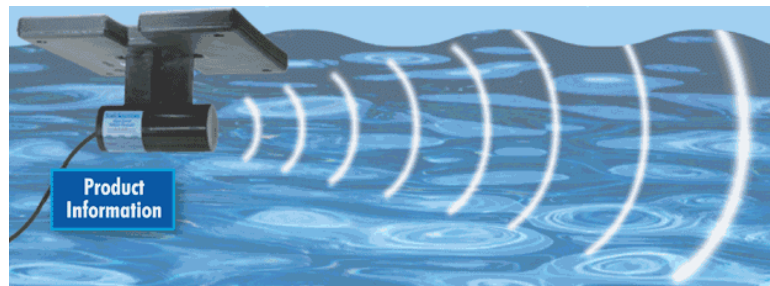
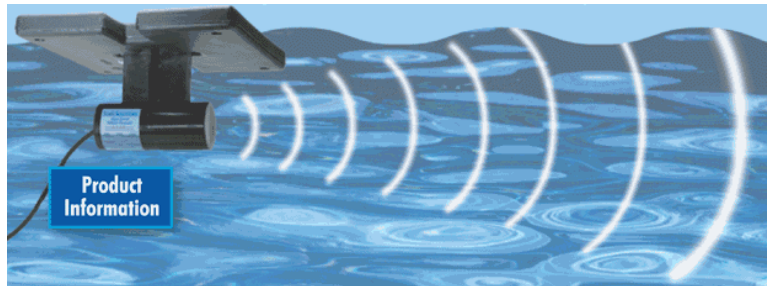
Planned Activities for Next Period

ASU

- Design PCR primers to specifically detect 16S rRNA of heterocystous cyanobacteria that are potential toxin-producers
- Continue efforts to screen for toxins from additional cyanobacteria obtained from Arizona Canal and Saguaro Lake
- Apply optimized PCR protocols to monitor potential toxin-producing cyanobacteria in the Arizona Canal and Saguaro Lake

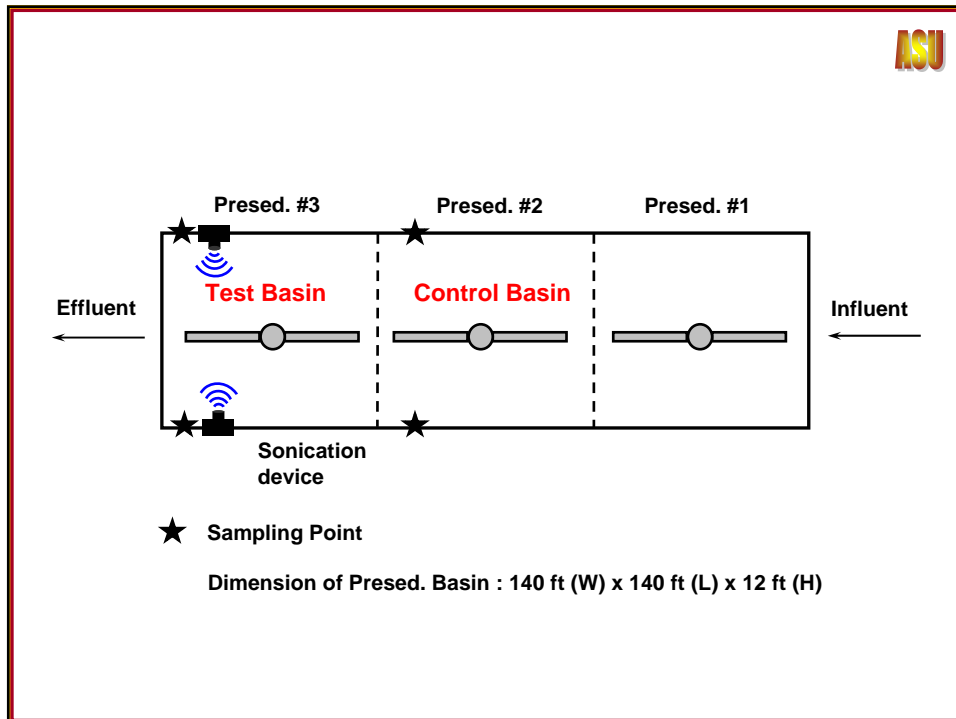
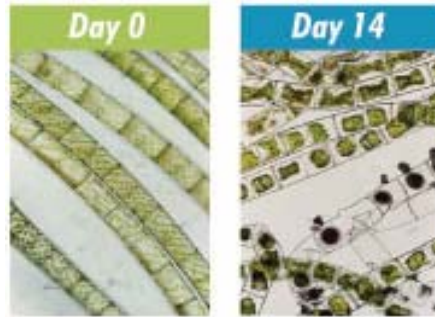
Sonication for algae control

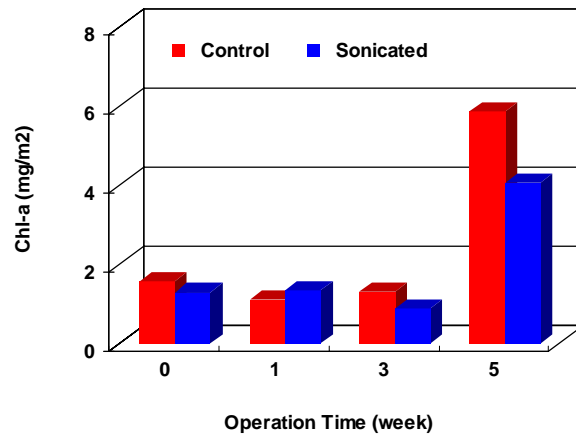
Youngll Kim



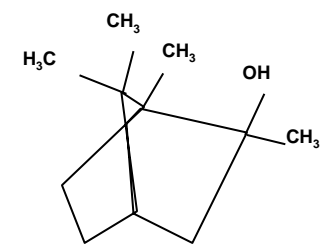
- ✓ **Ultrasonic algae control device is the state-of-the-art, gets rid of algae without harming other aquatic life**
- ✓ **Ultrasonic device is environmentally friendly, easy to use, cost-effective and uses no chemicals**

- ✓ Ultrasonic device generate ultrasonic waves that shock and kill the algae by ripping the vacuole of the algae cell

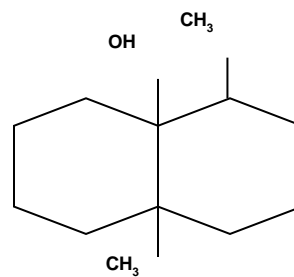




Wrapping Up



2-Methylisoborneol (MIB)



Geosmin



OTHER ACTIVITIES IN PAST YEAR

- ◆ Continued to publish/distribute water quality newsletter
- ◆ Collected extra samples for Phoenix and ASU from Roosevelt Lake to understand DOC runoff issues
- ◆ Cultured DOC-producing algae to learn more about their contributions to nitrogen containing DBPs
- ◆ Collaborated with S. Nevada Water Authority to screen isolated cyanobacteria for toxins and to improve analytical capability & understand reasons for fish kills in Saguaro Lake
- ◆ Completed evaluation of biocide coatings for canal linings
- ◆ Evaluate new technologies (sonic) and products (GAC) to improve water quality treatment



FUTURE DIRECTIONS/ACTIVITIES

- ◆ Arizona Virtual Water University
- ◆ AwwaRF Proposal
- ◆ Facility For Toxin Analysis
- ◆ Portable MIB sensor
- ◆ Other Ideas?



ARIZONA VIRTUAL WATER UNIVERSITY

- ◆ **“In 2006, we will establish a virtual water university that unites the cutting edge work in each university is doing on water management into one supercenter of research, community assistance and economic development.”**

Governor Napolitano, Arizona Town Hall at the Grand Canyon



ARIZONA VIRTUAL WATER UNIVERSITY

MISSION:

- ◆ **Serve as hub of research and technology development to give Arizona the tools to assure clean and sustainable water resources for the next century;**
- ◆ **Provide education, information, and analytical support to the public, government decision makers, water professionals, industry, and others about using, conserving, and managing water in arid environments;**
- ◆ **Be a resource for new water management technologies that produce new products and services for Arizona companies to export worldwide, thus creating a major new economic driver for Arizona.**



AwwaRF PROPOSAL

“STRATEGIES FOR CONTROLLING AND MITIGATING ALGAL GROWTH WITHIN WATER TREATMENT PLANTS”

In response to RFP 3111

**Collaboration between Malcolm Pirnie, Inc., ASU
and
14 Water Treatment Plants/Districts**



PARTICIPATING WATER TREATMENT PLANTS

◆ Chandler	AZ
◆ Peoria	
◆ Phoenix	
◆ Alameda CWD	CA
◆ Contra Costa WD	
◆ Santa Clara	
◆ Denver Water	CO
◆ Tampa Bay W	FL
◆ Central Lake CJWA	IL
◆ Indianapolis W	IN
◆ Minneapolis WW	MN
◆ St. Paul RWS	
◆ Philadelphia WD	PA
◆ Greenville	SC
◆ Newport News WW	VA

OBJECTIVES

- ◆ Collect and analyze existing information on types of algae found in water treatment plants
- ◆ Document the dominant algal types found in water treatment plants
- ◆ Identify algal issues triggered by modifications of treatment trains
- ◆ Develop case studies of treatment plants that are controlling/mitigating algae using different strategies
- ◆ Develop recommendations and guidance for utilities on
 - sampling and analysis to address algal issues within the plant
 - optimal control strategies that work for other utilities
 - best practices for operation and maintenance to reduce algal problems in treatment plants

ADVANCED WATER QUALITY EVALUATION AND MANAGEMENT STRATEGY

GOAL:

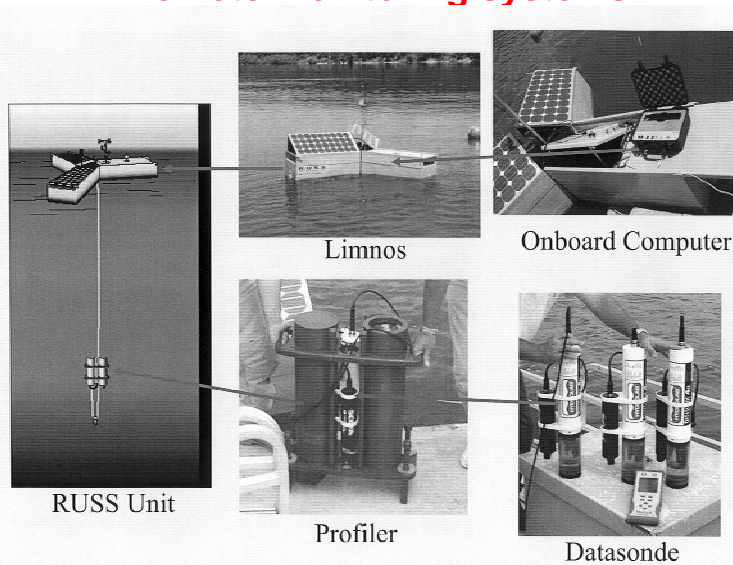
**TO RESPOND TO THE RECENT AND
EMERGING CONCERNS ABOUT TOXIC ALGAE**

RESOURCE FOR TOXIC ALGAE AND TOXIN ANALYSIS

Establish an ASU facility with the capabilities to:

- ◆ Develop PCR primers specific for potential toxin-producing algae
- ◆ Develop real-time PCR protocol for quantitative detection of specific potentially toxic algae
- ◆ Apply these methods to detect the occurrence of potentially toxic strains
- ◆ Isolate and characterize potential toxic strains
- ◆ Develop an optimized HPLC/MS protocol for rapid toxin analysis

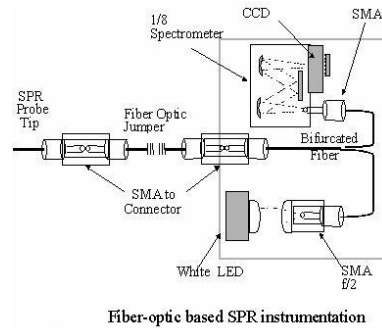
Continue to Urge SRP & CAP to install remote monitoring systems



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In-situ MIB Sensors

- Goal: use biomedical type sensors based upon surface plasma resonance technology to develop a field portable MIB sensor
- Initial work would start in 2006 and then external funding would be sought
- Inexpensive MIB sensor could be used to optimize WTP processes, find hot-spots of MIB production in canals, rivers, and lakes, and be used to assess customer complaints



Future directions & discussion

What do you see as the biggest challenge for the fall and into 2006?

What research would you like to see expanded?



Visit Our Websites

<http://ceaspub.eas.asu.edu/pwest/tasteandodor.htm>

Arizona Canal – Water Temperatures

